

S rep om ce

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ABSTRACT

Genome sequencing projects in the last decade revealed numerous cryptic biosynthetic pathways for unknown secondary metabolites in microbes, revitalizing drug discovery from microbial metabolites by approaches called genome mining. In this work, we developed a heterologous expression and functional screening approach for genome mining from genomic bacterial artificial chromosome (BAC) libraries in *Streptomyces* spp. We demonstrate mining from a strain of *Streptomyces rochei*, which is known to produce streptothricins and borrelidin, by expressing its BAC library in the surrogate host *Streptomyces lividans* SBT5, and screening for antimicrobial activity. In addition to the successful capture of the streptothrin and borrelidin biosynthetic gene clusters, we discovered two novel linear lipopeptides and their corresponding biosynthetic gene cluster, as well as a novel cryptic gene cluster for an unknown antibiotic from *S. rochei*. This high-throughput functional genome mining approach can be easily applied to other streptomycetes, and it is very suitable for the large-scale screening of genomic BAC libraries for bioactive natural products and the corresponding biosynthetic pathways.

IMPORTANCE

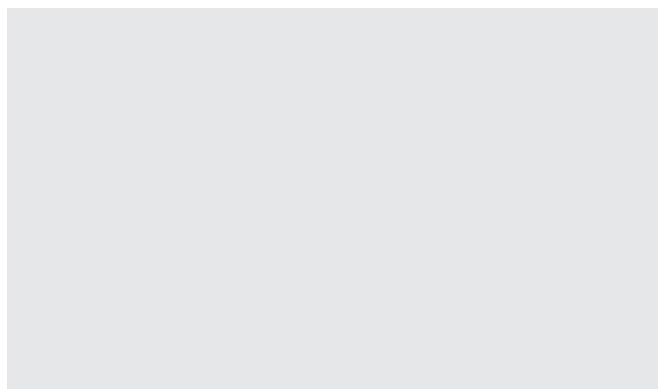
Microbial genomes encode numerous cryptic biosynthetic gene clusters for unknown small metabolites with potential biological activities. Several genome mining approaches have been developed to activate and bring these cryptic metabolites to biological tests for future drug discovery. Previous sequence-guided procedures relied on bioinformatic analysis to predict potentially interesting biosynthetic gene clusters. In this study, we describe an efficient approach based on heterologous expression and functional screening of a whole-genome library for the mining of bioactive metabolites from *Streptomyces*. The usefulness of this function-driven approach was demonstrated by the capture of four large biosynthetic gene clusters for metabolites of various chemical types, including streptothricins, borrelidin, two novel lipopeptides, and one unknown antibiotic from *Streptomyces rochei* Sal35. The transfer, expression, and screening of the library were all performed in a high-throughput way, so that this approach is scalable and adaptable to industrial automation for next-generation antibiotic discovery.

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1 3.5 μ, 2.1 150 () 2 (0.05%) 0.05% 0 0% 10%, 3 75% 25% 12 70% 30%, 16 1 8 100%, 1 8 25 0% 10% 0.3 / (30).

METHODS AND MATERIALS

Bacterial strains, plasmids, and culture conditions.

35 (7007) 5 Δ Δ KLΔ 3- LI3600 L(88) (20)

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Metabolic analysis of *S. rochei* Sal35 and BAC-containing *Streptomyces* strains.

35 (35) 6 30 35 3 1152/ 81 1154/ 86 3 1152/ 81, 1154/ 81, 1 8 81 35 20 5 () 210 () 40 (20 μ) ().

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Construction of the genomic BAC library of *S. rochei* Sal35.

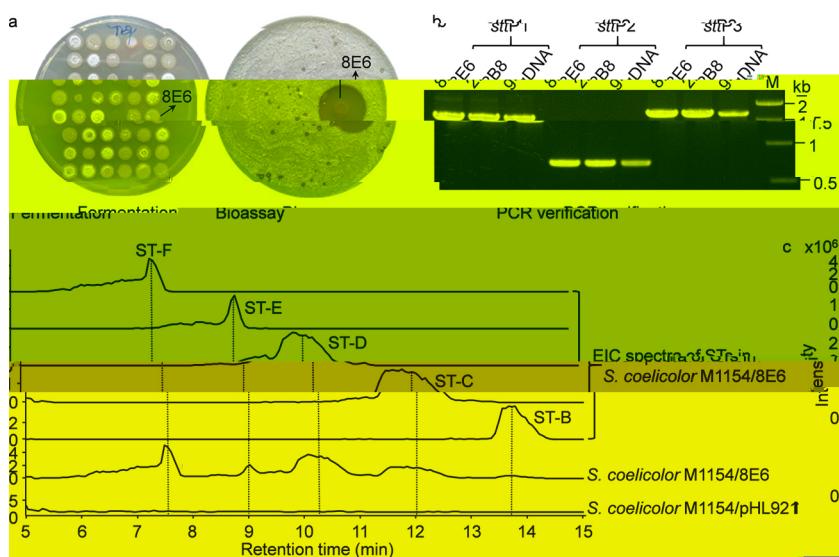
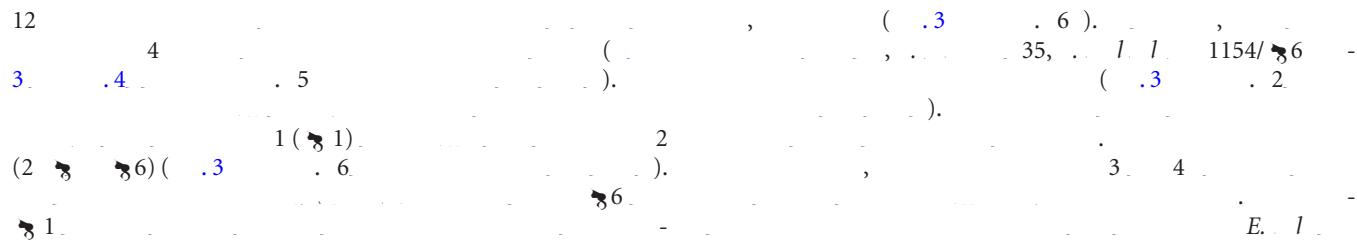
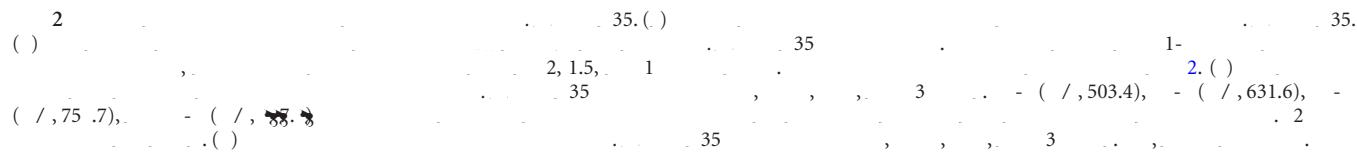
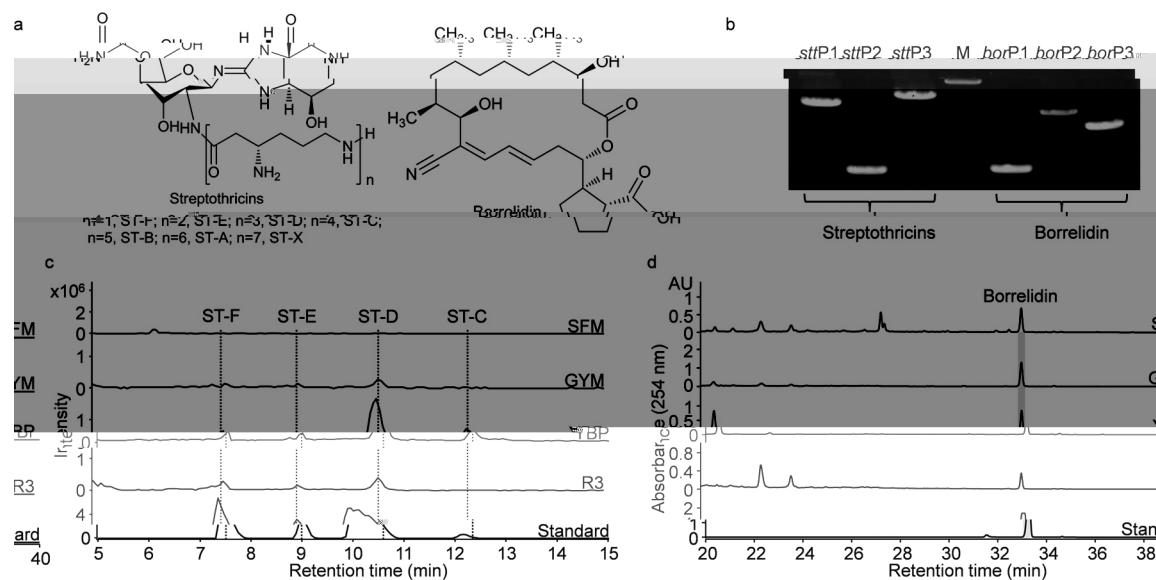
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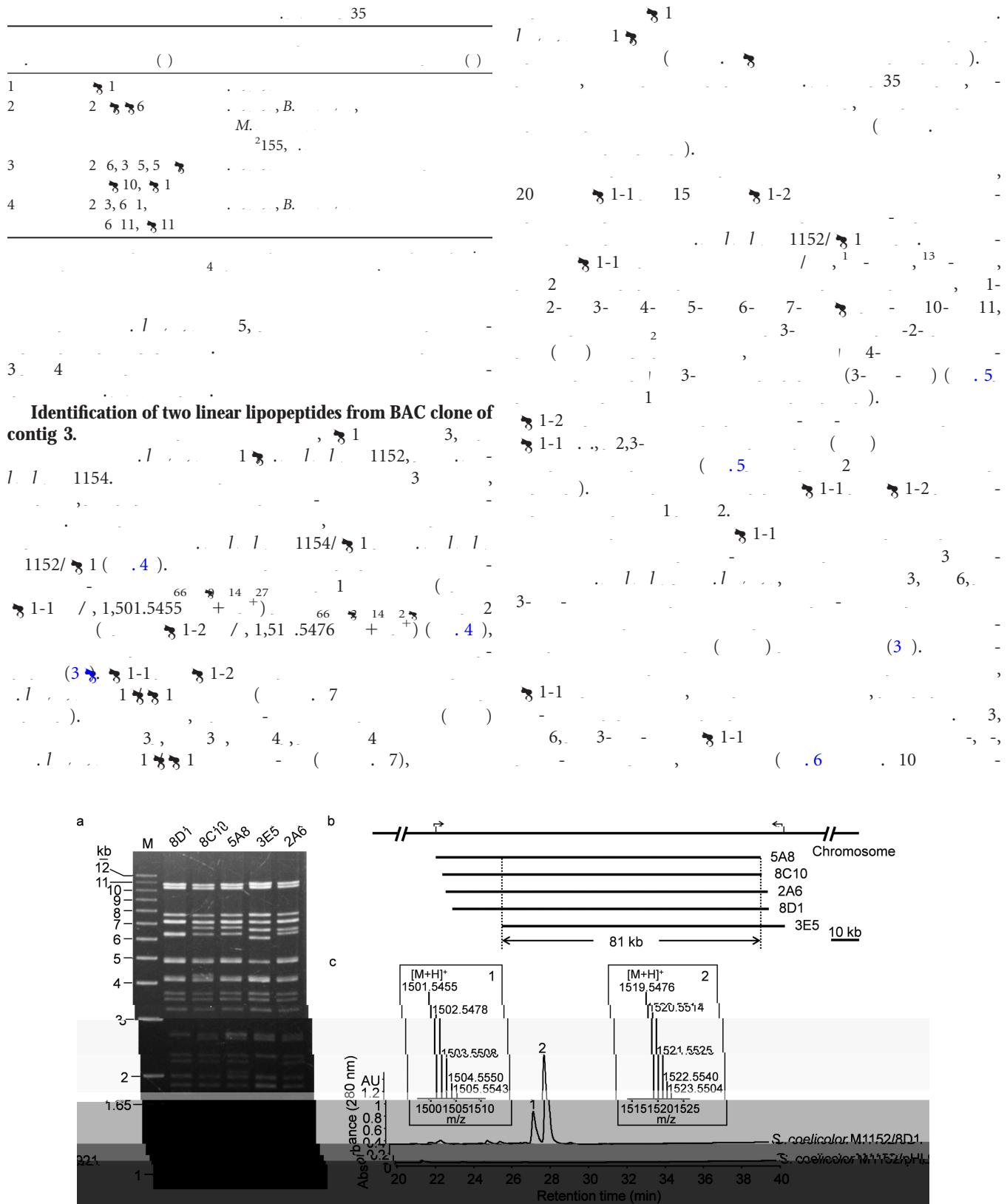
JEXAS screening of the *S. rochei* Sal35 BAC genomic library

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Production of streptothricins and borrelidin by *S. rochei* Sal35 and the LEXAS clones.

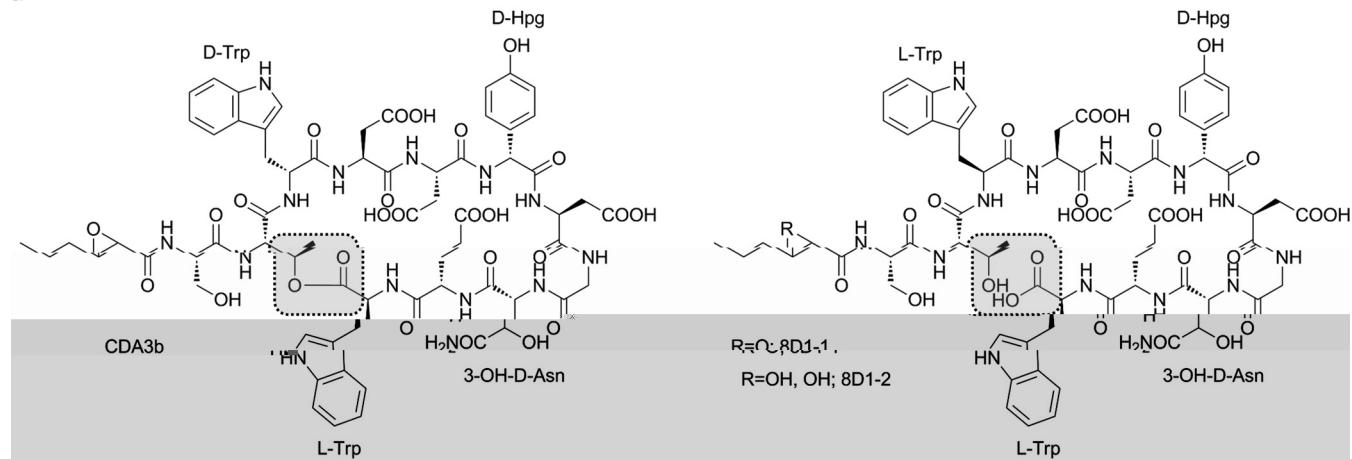


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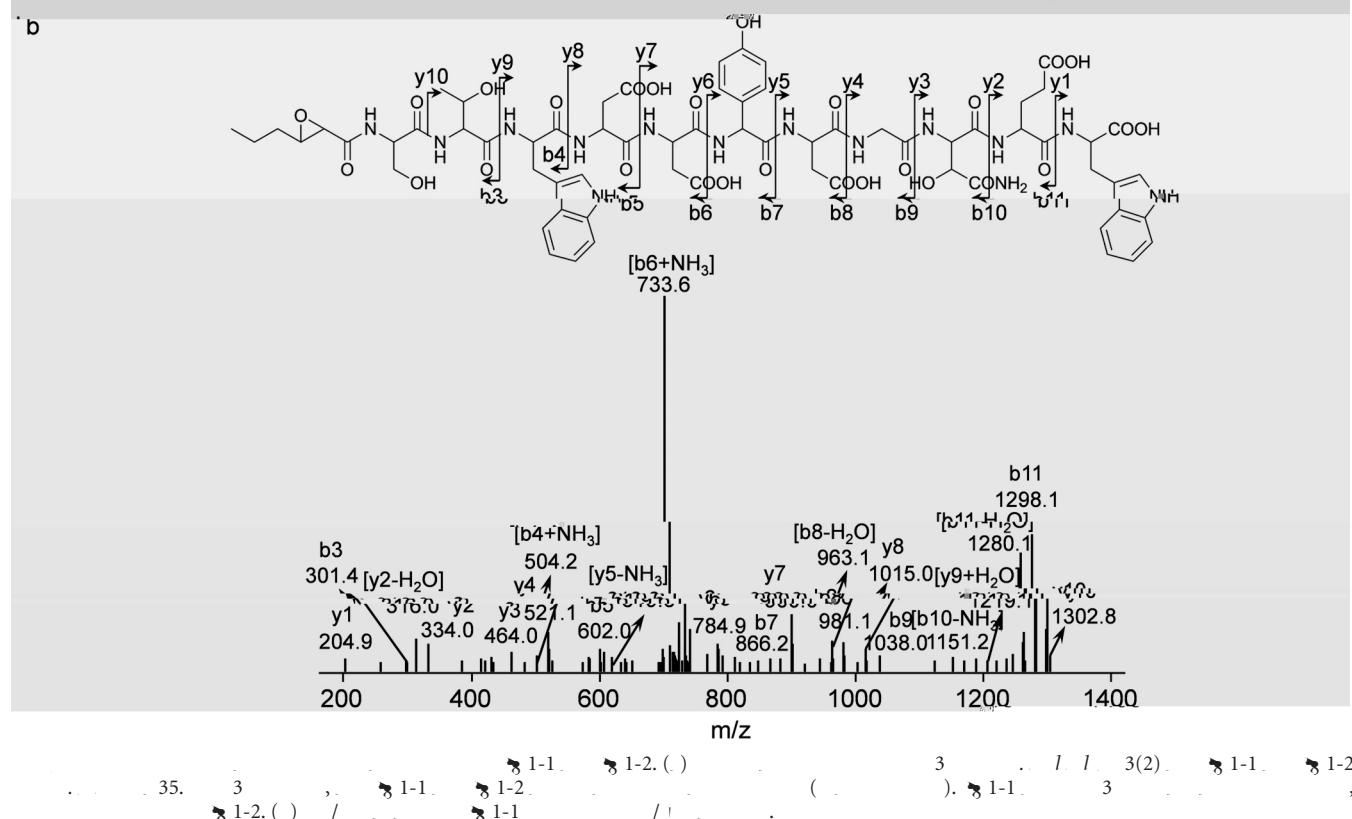


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a



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Bioinformatic analysis of the *cda2* gene cluster from *S. rochei* Sal35 for the biosynthesis of 8D1-1/2.

ACKNOWLEDGMENTS

(73 200 11 05 63 2010 10 201),
(31370134), (0 077 533-0 001) ,
(15 1400401).

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(31370134),
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(-0 3077 3533-0 001), () (15, 1400401), ()
(200 11 305 2010 10 201).

REFERENCES

1. , . 2012. 30 1 2010. 311 335. //
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