

Tianning Pu¹, Zhiling Mei², Wei Zhang¹, Wei-Jun Liang³, Xiufen Zhou¹, Jingdan Liang^{1*}, Zixin Deng^{1*}, and Zhijun Wang^{1*}

1, State Key Laboratory of Microbial Metabolism, School of Life Science and Biotechnology,
Shanghai Jiao Tong University, Shanghai, People's Republic of China.

2, Shanghai Thinkgene Biotech CO., LTD, Shanghai, People's Republic of China.

3, Department of Life and Environmental Sciences, Faculty of Science and Technology,
Bournemouth University, Talbot Campus, Fern Barrow, Poole, Dorset, United Kingdom, BH12
5BB

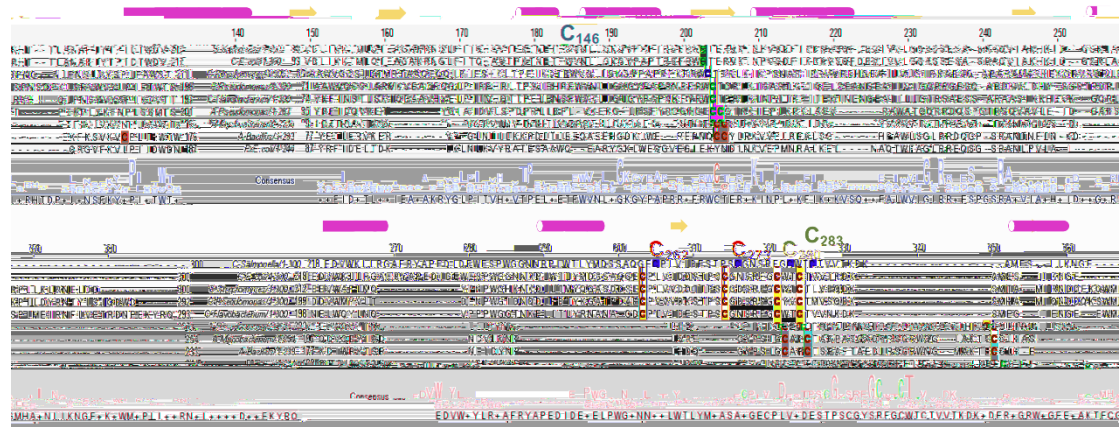
Running title: DNA Phosphorothioate Modification Reaction

* Corresponding authors:

Zhijun Wang, Ph.D., Laboratory of Microbial Metabolism and School of Life Sciences & Biotechnology, Shanghai Jiao Tong University, Shanghai 200030, CHINA, Tel/Fax: +86 21 62933765-2061(O), E.mail: wangzhijun@sjtu.edu.cn.

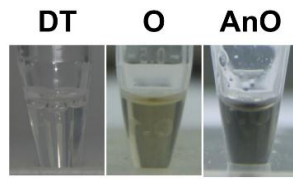
Jingdan Liang, Ph.D., Laboratory of Microbial Metabolism and School of Life Sciences & Biotechnology, Shanghai Jiao Tong University, Shanghai 200030, CHINA, Tel/Fax: +86 21 62933765-2061(O), E.mail: jdliang@sjtu.edu.cn.

Zixin Deng, Ph.D., Laboratory of Microbial Metabolism and School of Life Sciences & Biotechnology, Shanghai Jiao Tong University, Shanghai 200030, CHINA, Tel/Fax: +86 21 62933765-2061(O), E.mail: zxdeng@sjtu.edu.cn.

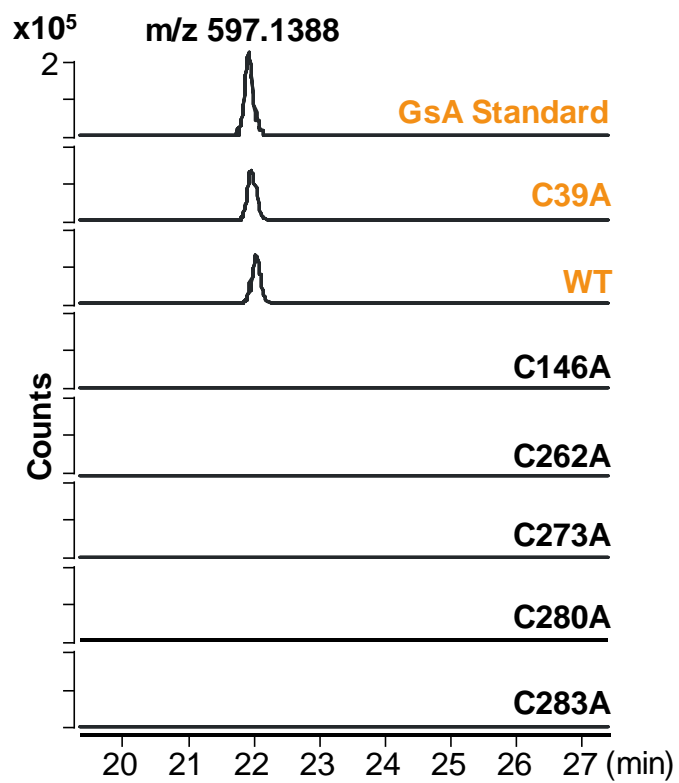


I c

Amino acid sequences of DndC from serovar Cerro 87, B7A, , , and APS reductases from , , and PAPS reductase from . were aligned using Jalview2.10.5 (Waterhouse , 2009). Residues from 93 to 300 are shown. Cysteine residues are highlighted with red color. Cysteine residue positions in the sequences of DndC from serovar Cerro 87 are labeled on the top of the residues. The residue C₁₄₆, C₂₈₀ and C₂₈₃ are involved in the coordination of Fe-S cluster (green color), while the residue C₂₆₂ and C₂₇₃ are not (red color). The secondary structure of DndC was predicted using PSIPRED Protein Analysis Workbench (McGuffin , 2000). The predicted α -helixes are shown as fuchsia roundrect, and the predicted β -strands are shown as saffron arrow. Numbers on the right show the actual length of the corresponding amino acid sequences.



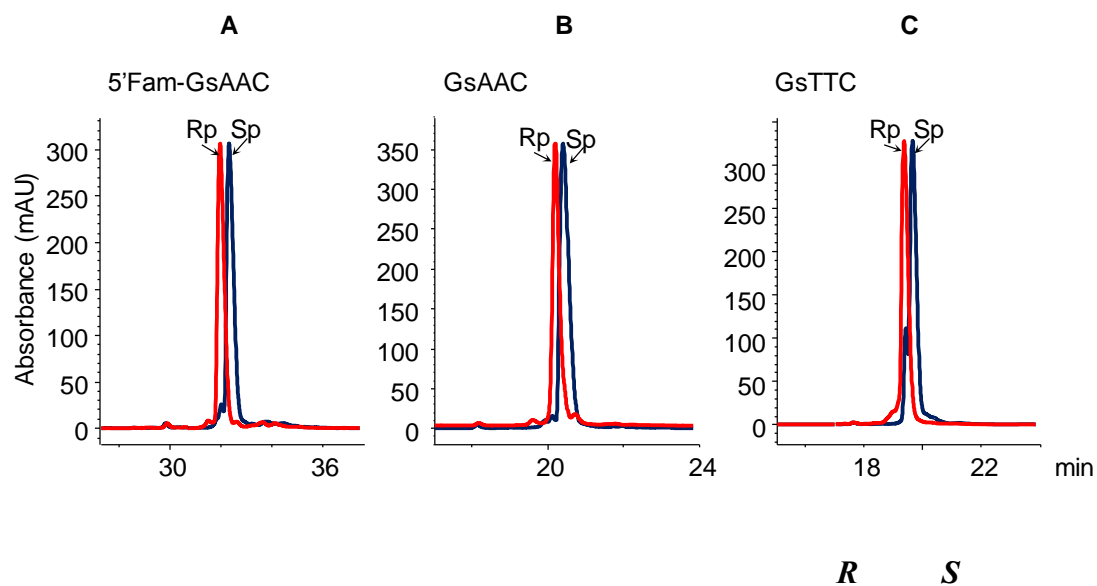
DndCDE was purified in the presence of oxygen as described in the Experimental Procedures section (indicated by 'O'). The protein was then treated using 1 mM 2,2'-dipyridyl ('DT'). The Fe-S cluster can be re-constituted under anoxic conditions using the 2,2'-dipyridyl treated DndCDE ('AnO').



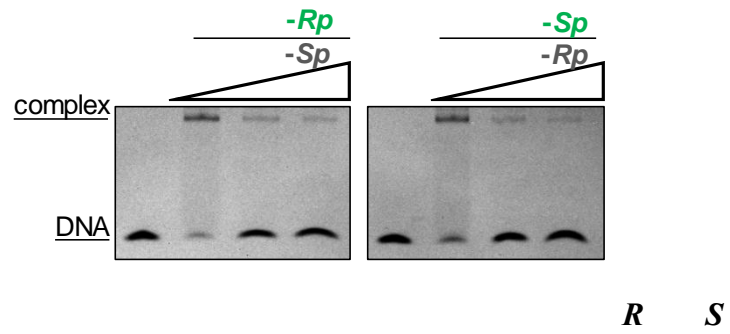
I

d d

The synthesized dGsA standard has a calculated m/z 597.1388.



Synthetic, commercially obtained ssDNA 24 oligonucleotides (DNA1/2 in Table 1) were separated, purified and detected by HPLC. Traces A, B and C show the (red) and (deep blue) configuration of PT oligonucleotides respectively. Table S1 lists the HPLC conditions for their separation.



Substrate DNA labeled with fluorescence is indicated by green color. Cold DNA is indicated with gray color. Ten or twenty folds more non labeled cold DNA was used for the experiments.

R S

Oligonucleotide	Gradient
24GsAAC (DNA1)	0-5 min, 60 % B*
Fam-24GsAAC (Fam-DNA1)	0-8 min, 70 %-75.2 % B, 8.1-9 min, 75.2 %-70 % B
24GsTTC (DNA2)	0-8 min, 57 %-61.2 % B, 8.1-9 min, 61.2 %-57 % B

B: solvent B, 10 mM Tris·HCl pH 8.0 and 1 M NaCl. Solvent A was 10 mM Tris·HCl pH 8.0.

References:

- McGuffin, L. J., Bryson, K., & Jones, D. T. (2000). The PSIPRED protein structure prediction server. *Bioinformatics*, 16(4), 404-405. doi:10.1093/bioinformatics/16.4.404
- Waterhouse, A. M., Procter, J. B., Martin, D. M., Clamp, M., & Barton, G. J. (2009). Jalview Version 2 - a multiple sequence alignment editor and analysis workbench. *Bioinformatics*, 25(9), 1189-1191. doi:10.1093/bioinformatics/btp033