RESEARCH PAPER



The Hippo/STE20 homolog SIK1 interacts with MOB1 to regulate cell proliferation and cell expansion in Arabidopsis

Jie Xiong, Xuefei Cui, Xiangrong Yuan, Xiulian Yu, Jialei Sun and Qingqiu Gong*

Tianjin Key Laboratory of Protein Sciences, Department of Plant Biology and Ecology, College of Life Sciences, Nankai University, Tianjin 300071, China

* Correspondence: gongq@nankai.edu.cn

Received 3 July 2015; Accepted 26 November 2015

Editor: James Murray, Cardiff University

Abstract

Multicellular organisms co-ordinate cell proliferation and cell expansion to maintain organ growth. In animals, the Hippo tumor suppressor pathway is a master regulator of organ size. Central to this pathway is a kinase cascade composed of Hippo and Warts, and their activating partners Salvador and Mob1/Mats. In plants, the Mob1/Mats homolog MOB1A has been characterized as a regulator of cell proliferation and sporogenesis. Nonetheless, no Hippo homologs have been identified. Here we show that the Arabidopsis serine/threonine kinase 1 (SIK1) is a Hippo homolog, and that it interacts with MOB1A to control organ size. SIK1 complements the function of yeast Ste20 in bud site selection and mitotic exit. The *sik1* null mutant is dwarf with reduced cell numbers, endoreduplication, and cell expansion. A yeast two-hybrid screen identified Mob1/Mats homologs MOB1A and MOB1B as SIK1-interacting partners. The interaction between SIK1 and MOB1 was found to be mediated by an N-terminal domain of SIK1 and was further confirmed by bimolecular fluorescence complementation. Interestingly, *sik1 mob1a* is arrested at the seedling stage, and overexpression of neither *SIK1* in *mob1a* nor *MOB1A* in *sik1* can rescue the dwarf phenotypes, suggesting that SIK1 and MOB1 may be components of a larger protein complex. Our results pave the way for constructing a complete Hippo pathway that controls organ growth in higher plants.

Key words: Arabidopsis thaliana, cell division, cell expansion, Hippo, MOB1, organ growth, SIK1, STE20.

Introduction

NRR ⊠t 📜 🕅 , 🕅 t 🕅 NY × × × ν T, (, t, 🖾 🖾 t, 🔟 , At, 🛛 t 🖾 🖾 🖉 -İQ, ,)Q)Q (, , KA KA Q Q KA 13 (t. - 🖾 🔤 t (🛛 XX X. t t t 🗱 🗱 🔣 t ₩<u>₩</u>₩, (K<u>₩</u>, <u>8</u>, 2009; G ₩ *e a*., 2012; M 🛚 L 🕅 🖾 . 2012: H Żt ⊠v ⊠y⊠yy L ⊠ , 2014). ⊠t-18 t 🕅 🕅 🕅 ⊠t⊠ _y, 🛛 🕅 (haa) , ((haa) haa haa haa haa haa haa haa haa G-(GRF) (a ., 2015), Reg a g Fac

TEQSINTEBRANCHEDI/CYCLOIDEA/PCF (TCP)(\blacksquare \blacksquare \bigcirc \bigcirc \bigcirc \frown
(D), e a., 2006), UBIQUITIN-SPECIFIC PROTEASE $\begin{array}{c} (\mathbf{B} \ \mathbf{B} \ \mathbf{S} \$ 2007; BX, (, e a., 2010; XKe a., 2013).

ଷାର୍ଥ୍ୟ ଅନ୍ମାନ୍ତ୍ର ମାନ୍ତ୍ର ମାନ୍ତ୍ର ମହା ଅନ୍ମାନ୍ତ୍ର ମହା 🕅 $\begin{array}{c} \underline{\ } \\ \underline{\ }$ 1⊠.t.bdd.

 $\begin{array}{c} & & & \\ & & & & \\ & & & \\ & & & & & \\ & & & & \\ & & & & \\ & & & & \\ & & & & \\ & & & & \\ & & & &$

 Image: Note: Image: KAR (a, t, have here a $\begin{array}{c} G_{1} \\ G_{2} \\ G_{3} \\ G_{4} \\ G_{5} \\ G_{7} , t_X (**b** e a , 2009).

 $\mathbf{B}_{\mathbf{A}} = \left[\mathbf{B}_{\mathbf{A}} + \mathbf{M} \right] \mathbf{M} = \left[\mathbf{M} \right] \mathbf{M} =$ Śa_n≴nowy now (nownow), (nownow), t, (21-__ now_t, (AK.) _t t haa -t aanana _a _ bhaa _ , j ta haa _t t haa C-t aanana $\begin{array}{c} \underline{\ } & \underline{\ } & AK-I & \underline{\ } & \underline{\ \ } & \underline{\ }$ Mana, yt. 🖄 , t. 🖄 Mana Marta, 18t, 📜 🔤 🖄 , 🕵 🖄 , 🖄 At \mathbb{A} t \mathbb{A} GCK [0] [0] [0] [0] \mathbb{A} \mathbb

38, _Σ F-κΒ ΝΑΤΑΣΔΑΤΑ _ t _ y ,ΝΔ _ t , ΝΔ , ΣΝΔ _ -

 $1 \qquad \qquad \mathbb{N} \qquad \mathbb$ e a = 2011). And $\mathbb{M} A \mathbb{M} \mathbb{M} \mathbb{M} MOBI$ t $\mathbb{M} \mathbb{M} OBI$

 $\begin{array}{c} \underline{M}t, \dot{H}M \\ H, \dot{M} \\ \end{array}, \begin{array}{c} \underline{L} \\ \underline{J} \\ \underline{J$ $\underline{\quad} E20 \underline{\quad} E20 \underline$ e a., 2002; K_X + e a., 2010). It. ____ K X A K + t , $\underline{1}$, \underline{1} , $\underline{1}$, \underline{1} , $\underline{1}$, \underline{1} , $\underline{1}$, $\underline{1}$, \underline{1} , $\underline{1}$, $\underline{1}$, \underline{1} , \underline{1} , $\underline{1}$, \underline{1} , \underline{1} , $\underline{1}$, \underline{1} , \underline{ $\begin{array}{c} \underline{\textbf{M}} \\ \underline{\textbf{M}}$ At B1B, IKINAMATE MANAGE A LAMA . NAME A -NAME A BAR A BAR A LAMA A A BAR Y Y MALE A MARY Y -t, 🖾 📶 (1) 🖞 🕅 K1. IN t 🖄 🕅 Ky, B1, IK1 ້ນໝະເຊັ້ນຊະຊຸເເເຊຍ , ນອະເພັ IK1 ເເເຊ, , _ຍ ເ, -ເຊຍນໝູ ເຊັ້ນໝະຍັ IK1_ ຊີເເ, ຊີ ຮ້າຍ ຍັນເ, ຊີ, _ຍ , ນອ້າຍຮັບເK1. ກາໝຍ_ y, ແລ້ນຊີ ຂັງ- $\begin{array}{c} & \underline{\textbf{M}} \textbf{K}, t, t, t, t, \textbf{I} \textbf{K} \textbf{I} \underline{\textbf{M}}, \textbf{K} \underline{\textbf{M}} \underline{\textbf{M}}, \textbf{K} \underline{\textbf{M}}, \textbf{K} \underline{\textbf{M}}, \textbf{K} \underline{\textbf{M}} \underline{\textbf{M}}, \textbf{K} \underline{\textbf{M}} \underline{\textbf{M}}, \textbf{M} \underline{\textbf{M}} \underline{\textbf$ _t _y 🖄 🏧 🕅 _121 t .

Materials and methods

Accession numbers

IK1, At1**&**69220; IA, At5**&**45550; IB, At4**&**19045.

Plant materials and growth conditions

1-1 (SALK 051369), 1-2 (SALK_010630), 1-3 $\begin{array}{c} (SALK_{046158}), & 1-4 \\ (SALK_{062070}), & 1-4 \\ (SALK_{0600}), & 1-4 \\ (SALK$ (SALK_046158), 1-4 (SAIL_636_C05, CS875528), b1a-1

Constructs and transgenic plants

Constructs and transgenic plants SIKI, t **booms** I **booms** G 🛛 🕅 t, C 🕅).

35S: YFP-M c-M bIA: M bIA, D A C - 12 M (- 10)Max (M - 10) (- $\begin{array}{c} \textbf{Max} \quad \textbf{Max} \quad \textbf{Max} \quad \textbf{Max} \quad \textbf{y}, \quad \textbf{y}, \quad \textbf{Max} \quad \textbf{Max} \quad \textbf{y}, \quad \textbf{Max} \quad$

Gene expression analysis

 $\begin{array}{c} \mathbf{A} \\ \mathbf{$

 $\begin{array}{c} & \overleftarrow{\textbf{b}} & \overleftarrow{\textbf{b}} & \overleftarrow{\textbf{b}} & \overleftarrow{\textbf{c}} & \overleftarrow{\textbf{c}} & \overleftarrow{\textbf{b}} & \overleftarrow{\textbf{c}} & \overleftarrow{\textbf{c}$

Yeast complementation, yeast two-hybrid screen, and verification

 verification

 ...
 y. _ t

 ...
 y. _ t

 8A60
 3::LEU2)

 ...
 \mathbb{Z}
 \mathbb{Z} \mathbb{Z}
 \mathbb{Z}

Quantification of cell size and numbers

Quantification of cell size and numbers F, \boxtimes I, $\sum_{i=1}^{n}$ I, \sum_{i E, 2010.

Measurement of nuclear DNA content of leaf cells by flow cytometry

FAC, AN III, yt K (BD, A).

1464 | Xiong et al.

Laser scanning confocal microscopy (LSCM)

Results

The Arabidopsis SIK1 encodes a STE20-like kinase

. M E20/H . . . 1 וׁ⊠. ANK AL (B_1, L, _ . BLA kaka katakan (_t, Aka_ha ha katakan (_t_-(AI 10) (I 20 (I 20) S. ce e ae (HL007C, AAA35039.1), H M t1 🛚 t2 🕺 🖌 H a e (AAA83254.1 _⊠ t 🚬 🛣 . ÌNZ , =58% ĨNZ , t 20; 51% _⊠ 68% ÌNZ , ÌNZ , ; 50% _⊠ t1; 🛛 44% 🖾 60% 🕅 , t2). 66% 🗖

ka than ka than han t ka ha ka ta han ka han ka han ı, ⊠v 🕅 🖓 🕅 🖓 🖓 🕺 🕺 🕺 🕺 🖓 👘 🖓 🖄 🕺 🖓 🖓 🖄 $\mathbb{W}_{\mathcal{A}}$, 2). \mathbb{N} t, t, \mathbb{N} , \mathbb{N} , , \mathbb{I} \mathbb{K} \mathbb{W} , , \mathbb{N} \mathbb{N} , \mathbb{N} , \mathbb{N} , \mathbb{N} , \mathbb{N} , \mathbb{N} _🕺 _ 🛚 K (CBI 🖾 K) 🏧 / K 🖾 CL 2689098, 🕅 🥵 1 C). 🎽 ._t, 14 .40 .11 10 10 ._ (18 t. .. 18 IK 11 10 _18 ×. K. K. t. 20. A K. K. K. (.e a., 2000; H 💆 🛛 _⊠, , Ň⊠, , 2002), e20∆, _⊠ 🕺 , 🕅 tynnam 🕵 _⊠nam . <u>1</u>87. <u>1</u>. (. 10), (NX 1D). , t 🛛 131 , () XXX, , 45.8%, _ N 🖉 NE COLUMNE LE DE LE DE LE COLUMNE DE LE DE LE COLUMNE DE L \mathbf{M} S e20: SIK1 124 (100 1D), 🔟 🛛 🛛 y 5.0% 🖞 🚬 t 🖾 🕅 _ 12 🖎 _ , . . 1000 _ , . . 12 _ . NALL L 124 (4.3%, =138). . . . [M] ty [M] [M] y [M] [M] $[L_1 + t_1]$ No the ha

SIK1 is expressed in mature tissues and is posttranscriptionally regulated



Fig. 1. Arabidopsis SIK1 is a Ste20 and Hippo homolog. (A) Schematic representation of the domain structure of Ste20 family proteins. The serine/ threonine protein kinase domain and the domains that mediate protein–protein interaction (PPI), including the Cdc42/Rac interactive binding (CRIB) and G beta-binding (GBB) motif of Ste20, and the Sav/Rassf/Hpo (SARAH) domain of Hippo and Mst1, are shown. No PPI domain can be identified in SIK1. (B) Alignment of the Ste20 family proteins. The kinase domains are highly conserved. D371 and D389 of SIK1 are predicted active sites (boxes). (C) Phylogenetic tree of SIK1 homologs from selected land plants, with metazoan and yeast Ste20 homologs as outliers. The tree is constructed with Clustal W2-generated multiple sequence alignment of SIK1 homologs (NCBI protein cluster CLSN2689098, protein kinase domain-containing protein) using the Neighbor–Joining method and plotted in Treeview. (D) SIK1 restores abnormal budding phenotypes in *ste20*Δ. Yeast cells from the background strain TN124 and *ste20*Δ complemented with SIK1 (*ste20*Δ/SIK1) are normal in cell shape and budding site selection. *ste20*Δ cells are irregular in shape with an abnormal budding pattern. Arrows indicate budding sites. Scale bar=5 μ m in (D). (This figure is available in colour at JXB online.)

🕺 t, 🔤 🖾 y 🖄 t 🏧 7-9 y-99 _ XX 📲 🚾 yı, t, t, i 🚾 t, 14-(_y- (__Mt, G_... (**N** 2G). IM 📲 🖄 t 🌶 🕅 🖄 t 🖄 🔪 ty, (🕅 🔟 t, ' 1 t KKG N. Y ⊠y t M (t BERK $(\mathbf{N} \times \mathbf{Z} \mathbf{I} \mathbf{K})$. $\mathbf{I} \times \mathbf{X}$, $\mathbf{M} \times \mathbf{L} \times \mathbf{I}$, . 🧸 🔍 🖄 🖉 - 🕅 🕅 🕅 y, ₩, ty, t ₩ _ Ŋ X t (₩ 2E), _M t_t, 🛛 🖓 🖉 🛴 $(\mathbf{N} \times \mathbf{2C}), \mathbf{t} \in \mathbf{N} \times \mathbf{N} \times \mathbf{N}$ 1,1, .

风服 化风 图 . . t 🚾 . . . 🔞 . yK , XXXXX SIK1:SIK1-GUS (SIK1:GUS). IN 10-(_y-_18t , G t BRA 🔄 🕐 t, t, t 🍽 🏧 (🖓 🚬 Ż), 🔟 . . , Qi, 🕅 (2 (**IVA** 2L, 🔞 🛛 🖞 🖄 🙀 🙀 🐴 🕺 t, 🖾 t 🗖 🗶 , 📆 🚬 🗃 Not by how 3) Mar to SIKI: GUS R Ż SĮĶI GUS., tim 🕅 ŠIKI GUS 1 ⊠ t**u⊠**t (t SIK1:GUS (**NK**, 2L), KAR NAKAR I 📜 NA I. 🛛 🗸 Ŕ t-t0_0,0000_0_0_0__ ໂໝີ້ນີ້, ÎK1 ⊠ ເໂໝຍ. 1⊠, ໄຊ ... ເ_ ເ, ໄຊ ໜີ ແັກ່ໜ _ M , 18 _ . . . **NR** $X \times 35S: GFP-SIK1$ X t, Α, , 🙀 , 🛛 t_⊠y में 🕵 , 4). (





Fig. 2. Developmental expression and post-translational regulation of SIK1. (A) Semi-quantitative RT-PCR analysis of *SIK1* transcript in different organs. *TIP4;1-like* used as internal control. (B–K) Histochemical staining of *pSIK1:GUS* T3 homozygous plants. SIK1 promoter activity is detected in (B) 3-dayold etiolated seedlings, (C) 3-day-old light-grown seedlings, (D) 7-day-old seedlings, (E) primary root of 7-day-old seedlings, (F) quiescent center of the root apical meristem, (G) 14-day-old seedlings, (H) 4-week-old plants, (I) inflorescence, (J) opened flowers, and (K) pollen grains. (L–S) Comparison between expression of *SIK1* transcriptional and translational fusions. GUS activity in *SIK1:GUS* transgenic lines is further restricted compared with *pSIK1:GUS* lines in (L–Q) 10-day-old seedlings and (R, S) rosette leaves of 4-week-old plants. The arrows indicate the quiescent center in (F), and stipules of leaves one and two in (M) and (O). Scale bars=500 μm in (B), (C), (J); 1 mm in (D), (N); 100 μm in (E), (O), (Q); 50 μm in (F); 1 cm in (G), (H), (S); 5 mm in (I). (This figure is available in colour at *JXB* online.)

Growth of sik1 mutants is retarded

A that AB C -DANK NAN NAN (, 🛛 🕅 e a., 2002; A 🕅 e a., 2003) (🏧 3A). F yk, M С 1-4 (SALK 051369), 1-1 (SALK_046158), 1-2 (SAIL_636_C05, CS875528) _X № 1-3 (SALK 010630), XX , XX ., , ⊠t 🖄_ _ M Ň t. t <u>t</u>t SIK1 🙀 A (124 3B). A 1-3 Ø, ty ty XXX 1X à ÌM383,∰23 t,,:153, _⊠(/ÌM3t, t,.) (**XX** 3C). **XX** 🖞 💢 t, 🕅 388 , (1915) 1038 , 1218 tz, (1, t, t, t, ..., 🖄 t, (1)32 Ż ts way wa 1-4.

- D 協議 協協、 泡 t、 , _ 、 _ 、 _ 、 _ 、 」 20 国、2020、 t、 、 国 t、 - D ANSE (2020日) - 「 、 」 2020、 、 国、ty、, _ (2010日) - 2030 、 _ 2030NSE UBQ10:GFP-SIK1 _ 異国 図 t、 , _ 1020、 、 」 2030 1-4 図 、 1120、

A 🚬 🚾 t 💭 🕷 🕂 🔂 🕅 🕅 🕅 🕅 🕅 🕅 (🎦 🕵 4A). SIKI 🛣 GÖLL _________, t, 🖄 . 🖄 1-4, t, , . 🖓 , 🐄 , 🔯 , 🔯 🖄 🖄 , __⊠/ İq t, 1⊠ (..., 1≅),1⊠t, (_1⊠), t_Ìv⊠Ìv⊠_,y_1⊠_t 🕅 ty 🛚 ty 🕻 🙀 , 🕅 🕅 🗶 . у.(.А Х(....) Ďt⊠ t, ¶⊈ ⊠ , tĽ, , 1 _12t. 14 t, 18 _ 1 M. Ì 🖞 t. 🎗 (t. 🎾 -ty 🧠 🖓 Ì 🕅 t 1787 🛛 t_1021 🖾 10880 ((**NK** 4B K; , 🙀 , 🛛 t_⊠y 🛛 🖾 🖾 🔣 t t t <u>,</u> 3), , . , ⊠ ty , 🖄 Maada 📜 🚬 🖓 y t 🚬 – D A Maa 🕅 🖓 🖓 🖓 🖓 1-4.

Cell number, cell size, and the ploidy level are reduced in sik1-4

N 🖄 t ً⊠ XX **,** ,⊠. ty , Ż ÌΝ ØØ. 88. .t 🔞 🗃 🚾 t 🗃 (A) 🖄 7- y-⊠t∖ Ż 🕺 🖄 _ tt, _ 🖄 🕅 4- . . 🕅 - . . MKK, t 1⊠t,_1⊠/t, ∖ t_ 🕺 🕅 y , 🔊 🕅 🖉 **لغ**) Ì. 1-4 M/ İ₫ţ 🛛 ty , 🕅 , 🔟 у. , ţ, Α (😰 Ø,



Fig. 3. The *sik1* null mutant has a dwarf phenotype. (A) *SIK1* gene structure and the T-DNA insertions. Exons, introns, and untranslated regions are represented by gray boxes, gray lines, and white boxes, respectively. Insertion positions of the T-DNA in the four alleles are shown. (B) *sik1-4* is a null allele. An mRNA fragment representing the kinase domain (KD; exons 2–12) can be detected in *sik1-1*, *sik1-2*, and *sik1-3*, but not in *sik1-4*. *sik1-3* is a knock-down allele, with the T-DNA inserted into the seventh intron. (C) Phenotypes of four alleles. The three null alleles are slow in growth compared with the wild type, with *sik1-4* having the most severe phenotypes. Diameter of pot=6.5 cm in (C). (This figure is available in colour at *JXB* online.)

 $\begin{array}{c} \left(\begin{array}{c} \mathbf{\dot{N}} \mathbf{\dot{M}} \mathbf{\dot{L}} \mathbf{\dot{L}} \mathbf{\dot{M}} \mathbf{\dot{M}} \mathbf{\dot{L}} \mathbf{\dot{L}} \mathbf{\dot{M}} \mathbf{\dot{M}} \mathbf{\dot{L}} \mathbf{\dot{L}} \mathbf{\dot{L}} \mathbf{\dot{L}} \mathbf{\dot{M}} \mathbf{\dot{L}}

, and a main a said a said a said a said a said a said a said a said a said a said a said a said a said a said t**ing** .1., KØ, _ NAR, _ NARK t, NA NAR, NA ty (, <mark>\$_y_, 2013). }}</mark> M \mathbb{N} \mathbb{N}_{t} 🛛 _Ż 2014), Y yt 🖞 tŻy _ℤ y 🖄 _ t , Ż, Ż , tt, , Ż. Ż4- , Ż- , _⊠t, t . Ì .`M2Kİt, 1%⊒ . 🛛 -🙀 🕅 🛛 💵 🔍 (, 1-4. _ _ , 🖄 🕅 y , , , 🐄 🖄 (🕅 t, 🕅 ty, (🏛 51).

t -,⊠`Nooka Na Na Na Na Na Na y t_ka __ , _t _₂ , ∆y , _∆y t_ka M İΧ XX <u>×</u> -12/12 v $\square \square _ t \square (ANT _ \square GRF5) (\square \square e a., 2005;$



Fig. 4. Complementation of *sik1-4* with *pUBQ10:GFP-SIK1*. (A) Semi-quantitative RT-PCR of *SIK1* in the wild type, *sik1-4*, and the complementation (Comp.) line. *TIP4;1-like* is used as internal control. (B) Seven-day-old, vertically grown seedlings. (C) Quantification of primary root length of seedlings at 7 days post-germination (dpg). (D) Four-week-old plants. (E) Maximum rosette radii of the wild type, *sik1-1*, and the complementation line, measured over a period of 5 weeks. (F) Eight-week-old plants. (G) Plant height measured over a period of 6 weeks. (H) Fully opened flowers. (I) Mature siliques. (J) Halves of siliques. (K) Dry seeds. Bars=SE in (C), (E), and (G). *** indicates *P*<0.001 (Student's *t*-test) in (C), (E), and (G). Scale bars=1 cm in (B), (I), 10 cm in (F), 1 mm in (H), (J), 200 μm in (K). (This figure is available in colour at *JXB* online.)



Fig. 5. sik1-4 has reduced cell numbers, cell sizes, and ploidy levels compared with the wild type (WT). (A) Root tips of 6-day-old vertically grown WT and sik1-4 seedlings. QC, guiescent center; MZ, meristematic zone; EZ, elongation zone. Arrows indicate the beginning of the MZ and EZ. (B) GUS staining showing pCYCB1;1 promoter activity in WT and sik1-4 root tips 7 days post-germination (dpg). (C) Root hairs (n >1000) of WT and sik1-4 seedlings at 7 dpg. (D) Distribution of root hair lengths in the WT and sik1-4. (E) The fifth rosette leaves of the WT and sik1-4 at 28 dpg. (F) Lower epidermis of (E) with representative cells highlighted. (G) Petals from fully opened flowers. (H) Lower epidermis of (G) with representative cells highlighted. (I) Flow cytometric analysis of the fifth rosette leaf of 28-day-old WT and sik1-4. A total of 20 000 nuclei are sorted for each sample. (J) Quantitative RT-PCR of core cell cycle marker genes and regulators from the first and second true leaves of 8-, 11-, and 14-day-old seedlings. CYCD3;1, G1 phase-specific marker; E2FD/DEL2, G1/S specific; CDC6, S-phase specific; CYCA2;1, S/G₂ specific; CYCB1;2 and CYCB2;1, G₂/M-phase specific markers. ANT and GRF5, transcription factors that regulate cell proliferation; KRP1, cell cycle inhibitor. Bars=SD. GAPC2 used as internal control. ***P<0.001, ** P<0.01, and *P<0.05 (Student's t-test) in (J). Scale bars=100 µm in (A), 50 µm in (B), (F); 1 mm in (C), (G); 1 cm in (E), 20 μ m in (H). (This figure is available in colour at *JXB* online.)

MOB1A and MOB1B were identified as SIK1interacting proteins

RÒRR B. Ì Ì BRÌR tÌ BR L, IKI İ B, Ì B B, _ 2H 🛛 🖾 M . **™**. **™**t_Ży ⊠ (_____2). And Make t 10, , 1A 1B (At4419045) (At5 45550) _ 212 , 1A ⊠ , 1B ίΩ (⊠ tÌ 🗱 B (Õ⊠t 🖾 e a., 2005, 2006), _⊠ _ t ⊠t, (, 1⊠, ty, , Ì N/ 2011; 🚾 _ e a., 2013), ____t_t_t_t_IK1, ___t, 1 ₩250 t, 🖄 t, _120 t, _t t, ₩250 t, ⊠-, Mat., Mat., t., t

SIK1 interacts with MOB1 at its N-terminal domain

IK1 _ , ,) (2000 ,) (2001 10 , 2000 , 200 , 201 () (2000 , 201) , 201 t) (201 2H. ⊠ İst, BD, 18tk, t. $\mathbf{M} (\mathbf{A} \in \mathbf{L}, \mathbf{A}), \mathbf{A} \in \mathbf{M} \in \mathbf{M} \in \mathbf{M} \times \mathbf{M} = \mathbf{M} (\mathbf{M} \in \mathbf{M} \times \mathbf{M}),$ \mathbf{X} L, $\mathbf{H}\mathbf{X}$ _t, 1- IK1 ($\mathbf{M}\mathbf{X}$ _ \mathbf{M} 1 235), M - IK1 (100 1 504), 1 - 1 K1 ___<u>`</u>A___ _ Mart, A_ Marke Mar . . t B1A _⊠∕ B1B (**DX** 6C). \mathbb{N} \mathbb{N} $\mathbb{L}_{\mathbf{x}}$ \mathbb{N} \mathbb{N} $\mathbb{A}_{\mathcal{L}}$ _t, 🛛 y - IK1 🖄 🖄 - 🏧 IK1 **B**1 B1 $\underline{\mathbb{M}}_{\underline{\mathbb{N}}}$, $\underline{\mathbb{N}}_{\underline{\mathbb{N}}}$ \mathbb{M}_{ℓ} t \mathbf{M} t Ececac (, \mathbf{M} \mathbf{M} \mathbf{M} 5), t \mathbf{M}

🕷 t 🗶 🕷 🛛 . t 🔍 🕷 🛛 IK1 🗖 B1A . 🕅 . _0 🕅 🕺 , . . 🛛 IK1, E F. _ , - 🕅 . B1A $[1, \underline{\mathbb{N}}] = t$, [1,t. B1A 🖻 E F . 🛛 (, t, t, (, . t 🔡 t.

 Table 1. sik1 has a lower cell number and reduced cell sizes compared with the wild type.

Parameter	Wild type (average ±SE)	sik1-4 (average ±SE)	Student's t-test
Length of RAM (μm), 7 dpg	296.6±4.2 (<i>n</i> =86)	182.8±2.5 (n=84)	<i>P</i> <1E-50
No. of protoderm cells in RAM, 7 dpg	38.2±0.4 (<i>n</i> =86)	24.0±0.3 (n=84)	P<1E-61
Length of RAM region with $pCYCB1$;1:GUS activity (μ m)	186.8±2.5 (<i>n</i> =108)	116.1±2.1 (<i>n</i> =73)	P<1E-51
Area of petal (mm ²)	1.85±0.03 (n=123)	1.00±0.02 (n=109)	<i>P</i> <1E-61
Area of petal epidermal cell (µm ²)	220.8±2.7 (n=100)	191.0±2.8 (<i>n</i> =86)	P<1E-12
Area of the fifth rosette leaf, 28 dpg (mm ²)	165.9±7.2 (<i>n</i> =28)	47.8±3.4 (n=28)	<i>P</i> <1E-17
Area of lower epidermal cell (µm ²)	8955.4±121.9 (n=420)	3271.5±37.3 (n=440)	<i>P</i> <1E-99
No. of palisade cells per 250 000 μm^2	93.5±3.0 (n=43)	210.2±6.7 (n=44)	<i>P</i> <1E-23

dpg, days post-germination.

 Image: Sector of the sector

Genetic analysis of SIK1 and MOB1A

Mar IK1 Mart 💆 t, 🕅 B1A, <u>R</u>, t. XXX XX X-, t 1αβ,t_1⊠t, _, `Nandana Manaza , 1⊠ ty , , Kajkat, ka_na_y, (t, , 🕂 🛯 🖉 🖉 🕼 🖉 🗸 t 🔍 🖾 SIKI 🖉 MOBIA. Ìn 🖾 t, 1- $4, _$ *bla-1* (GABL 719G04) (123 _ *e a* ., 2013) (123 $\dot{\mathbf{D}}$ $\dot{\mathbf{D}$ $\dot{\mathbf{D}}$ $\dot{\mathbf{D}}$ $\dot{\mathbf{D}}$ $\dot{\mathbf{D}}$ $\dot{\mathbf{D}}$ $\dot{\mathbf{D$ ͺϪ, ΣΣΜΑΣ, (___1 / bla/, yμαΣΕΝΑΣ C ((_t_Σ`t ...Σ)...,Σ, ΒΙΑ-, Σ, Σ, Σ, ΣΣΣΕΛΑΣ, (ΒΙΑ-Ε) 1\$\$\$, t_1⊠t_{*}, \$\$\$, , **`**\$\$\$, **x**(**b**\$\$\$, y(**b\$\$**\$\$, 4; , ..., \$\$\$, 18t_8y **b\$\$\$**\$, 6). , E $\mathbf{M}_{\mathbf{M}} = \begin{bmatrix} \mathbf{M}_{\mathbf{M}} & \mathbf{M}_{\mathbf{M}} \\ \mathbf{M}_{\mathbf{M}} & \mathbf{M}_{\mathbf{M}} \end{bmatrix} \begin{bmatrix} \mathbf{M}_{\mathbf{M}} & \mathbf{M}_{\mathbf{M}} \\ \mathbf{M}_{\mathbf{M}} & \mathbf{M}_{\mathbf{M}} \end{bmatrix} \begin{bmatrix} \mathbf{M}_{\mathbf{M}} & \mathbf{M}_{\mathbf{M}} \\ \mathbf{M}_{\mathbf{M}} \end{bmatrix} \mathbf{y}, \mathbf{t}$, 1886), 🖄 🖄 🖉 🖉 🖄 🕺 🖄 🕺 🕅 ., (🛛 , , , t , , , 🕅 🕅 İdit, t, 囟(內國 8C, G). , 囟, 內國人 1/ B1A-E, _ . [M] ty [M] [M] t I_{1} [M] , bIa/ [K1-E. [M] ty, ba, bba, t b1a. , ba, ba ba ba ba ba∭ո_⊠t t, 15վ,t_ _⊠ Hn⊠ . _t _y, t, ⊠, _y,⊠ _⊠ Ռոֆավ, ty B1A = [3] (M) = t (I = M) = [3] (M1888 1888 IK1- <u>1</u>84

Discussion

The molecular function of Hippo is probably conserved among eukaryotes

 $\begin{array}{c} & & \\ & &$

`¤_t_y._,.,™¤<u>k</u>k_v... to wave wave _t _y t_t, ⊠to +a, t_ _n ave a to t (_⊠, 2010; ___⊠ G, _⊠, 2013). ⊠, ⊠ E20-Wat Man Ha R K Ma y K Lt , 🖄 🚾 🖾 🖾 🙀 t, _ . t, 🚾 (, _ _t 💆 C, , 🚾 🕺 2015). $(\mathbf{X} \otimes \mathbf{X} \otimes$ 2006) _⊠ $\begin{array}{c|c} \textbf{M} & \textbf{M} & \textbf{M} & \textbf{M} \\ \hline \textbf{M} & \textbf{M} & \textbf{M} & \textbf{M} \\ \hline \textbf{M} & \textbf{M} & \textbf{M} & \textbf{M} \\ \hline \textbf{M} & \textbf{M} & \textbf{M} & \textbf{M} \\ \hline \textbf{M} & \textbf{M} & \textbf{M} & \textbf{M} \\ \hline \textbf{M} & \textbf{M} & \textbf{M} & \textbf{M} \\ \hline \textbf{M} & \textbf{M} & \textbf{M} \\ \hline \textbf{M} & \textbf{M} & \textbf{M} \\ \hline \textbf{M} & \textbf{M} & \textbf{M} \\ \hline \textbf{M} & \textbf{M} & \textbf{M} \\ \hline \textbf{M} & \textbf{M} & \textbf{M} \\ \hline \textbf{M} & \textbf{M} & \textbf{M} \\ \hline \textbf{M} & \textbf{M} & \textbf{M} \\ \hline \textbf{M} & \textbf{M} & \textbf{M} \\ \hline \textbf{M} & \textbf{M} & \textbf{M} \\ \hline \textbf{M} & \textbf{M} & \textbf{M} \\ \hline \textbf{M} & \textbf{M} \\ \hline \textbf{M} & \textbf{M} \\ \hline \textbf{M} & \textbf{M} \\ \hline \textbf{M} & \textbf{M} \\ \hline \textbf{M} & \textbf{M} \\ \hline \textbf{M} & \textbf{M} \\ \hline \textbf{M} & \textbf{M} \\ \hline \textbf{M} & \textbf{M} \\ \hline \textbf{M} & \textbf{M} \\ \hline \textbf{M} & \textbf{M} \\ \hline \textbf{M} & \textbf{M} \\ \hline \textbf{M} & \textbf{M} \\ \hline \textbf{M} & \textbf{M} \\ \hline \textbf{M} & \textbf{M} \\ \hline \textbf{M} \hline \hline \textbf{M} \\ \hline \textbf{M} \hline \textbf{M} \\ \hline \textbf{M} \hline \textbf{M} \\ \hline \textbf{M} \hline \textbf{M} \\ \hline \textbf{M} \hline \hline \textbf{M} \hline \hline \textbf{M} \hline \hline \textbf{M} \\ \hline \textbf{M} \hline \hline \textbf{M} \hline \hline \textbf{M} \hline \hline \textbf{M} \hline \textbf{M} \hline$ 2014). All t X MAL, t t E20 Éonna an the second se $\frac{1}{10}$, $\frac{1}$ ⊠,...⊠Ç,150⊠ 1508. 18 t,.., _2008,..., t,yt, , 1608..., $\mathbf{M} = \mathbf{M} =$ $A_t = \frac{1}{2} \begin{bmatrix} t & A & A \\ - & A & -A \\ -$ 🛚 t 🖼 🔤 t _t , IK1. _ _🕅 NALAN NALAN IN $\dot{\mathbf{M}}_{\mathbf{z}}$ t, t, $\mathbf{M}_{\mathbf{z}}$, $\mathbf{M}_{\mathbf{z}}$, t $\mathbf{M}_{\mathbf{z}}$, $\mathbf{M}_{\mathbf{z}}$ 💯 🔯 tj., 🖾 ty., 🔄 🖉 🖄 🖄 🖓 🖓 🕺 $(H_{\boxtimes}, y e a., 2003; M_{e} e a., 2003; M_{e} a., 2003; M_{e$ 2003; $(\underline{N} e \ a., 2003; \ e \ a., 2003)$. ÌMANARÝ, KALELI, IK1_, LA L, A ÌMAN KALINAN, Y IMANARI, V, NARINAN, KALINAN, MALANA, IMANAN, Y 🛛 y 🖾, 👖 🕅 ARE BINA NA (tashara (y) Binaka Da Bara (,) Da y) , 18 t, , , _18 108 18;, , 18 , t, _18 , 14 , 19 11 10 18 18 , t 1088 _1943, _1181, 1182 , 143,143 _183, t, 18,118,1182 [18,1182] 1183 1181 1182 1183 1181 1183 1181 1183 1181 1183

A possible structural basis for the kinase–scaffold interaction

 $t _ t _ _ _$ \mathbb{R}

1470 | Xiong *et al.*



Fig. 6. The N-terminal domain of SIK1 (N1-SIK1) is responsible for its interaction with MOB1s. (A) SIK1 is subdivided into four fragments for yeast two-hybrid (Y2H) analysis. (B) SIK1 and MOB1s do not have self-activation activity. (C) Interaction between the full length and fragments of SIK1 (plus the activation domain, AD) and MOB1s (plus the binding domain, BD) verified on triple- and quadruple-dropout plates. + and –, positive and negative controls. (D) Interactions between AD-MOB1s and BD-SIK1. (E) Interaction between SIK1 and MOB1A confirmed in tobacco leaf epidermal cells with bimolecular fluorescence complementation (BiFC). Scale bar=50 μ m in (E). (This figure is available in colour at *JXB* online.)





Fig. 7. Subcellular localization patterns of SIK1 and MOB1A. (A) RFP–SIK1 is co-expressed with GFP-tagged organelle markers. SIK1 is co-localized with *trans*-Golgi-network (TGN)/early endosome (EE) and plasma membrane (PM) markers. (B) RFP–MOB1A localizes to the nucleus (N), and co-localizes with TGN/EE, tonoplast, and PM markers. (C) When co-expressed with MOB1A, SIK1 can be detected in the nucleus (N). (D) N1-SIK1 localizes to the nucleus, and co-expression with N1-SIK1 brings SIK1 into the nucleus. Scale bars=25 μm in (A–D).

(, t<u>1</u> 🗰 🛛 e . a . , 2003; . . . -t 🖾 🕅 **⊠**, **⊠** *e a* ., 2004). - 🛛 🕅 t ⊠_, ∖⊠ (H2)_⊠ ₩ (L1) 💆. , 1 A 🖄 🖾 , , **) 🛛 🛍 🛄 🖾** y , , 🛍 , 🖄 🕻 (. 🗖 M . t**`)⊠ 10 1080 t**. . , . 10 . 10 . (10 . **K_10**80 . y, **. _\$\$**\$\$\$ 📜 🕅 🔟 , , , , _ _ E51 💭 E55, 🗓 , , , 🖾 , j 🖾 t. _, , 1A 🕅 🕅 🕅 🕅 (t. 1 🚾 🖉 e. a., 2003). C. 🖬 🛛 t 🛚 t y, t y,_tMMML, t_t_XMML, ™t10MML, ↓ t10ML 1. 🛣 _ D, 🕺 _N/ C, 🖄 1 🕅 🖏 , , () 🕅 🕵 y , _t_1. 2 2 2 🛛 🖾 У. _ t, _ . _ . , _ 🕺 () 🖓 t, 🖄 🕅 🕅 İğİQL, İQ∠, T⊠(, T⊠, T⊠ tİQ_t(t.) . 🛛 🖾 t XX. 1 🛛 🛛 📜 🖓 🖄 🕺 🕹 🗤 1 🔤 t . 🛛 🖾 🖉 🖉 . -t, 🔯 🏧 **D** $\mathbf{M} \mathbf{M} = (\mathbf{M}, \mathbf{M}, 1 🛛 \boxtimes , ₩ y_t t . ⊠ ⊠ () N2(, 1A, ,) N2 y_tt) N23 -t . ⊠, ⊠ e a., 2004), N2(t) N23 51 _⊠/ 55 _‡\$N233 -t. 🖾 🖾 🛛 · NARKAR, t_KNA _ NA _ NAR. × NØ, . 1A. × × × 🔟 🖾 🔟 🚬 . H 🖾 🚬 IK1, 🔟 B1 🛱 _y _ Mat 🛛 t t⊠ t_N⊠_ y NQ, t N200, ._⊠K.∉ XK.

How SIK1 may participate in organ size control

t 🕅 🛛 🖾 . (**t _t t** ,) - 1 b1150,t_120t. A thronistic y hashed a ty , M .,⊠N⊠t,⊠t N⊠t t , k⊈_tt__tt, t . . 🛛 t 🕅 🚺 🚬 🚬 t 🕅 🕅 משׁל מאשל (אם מי אם מי געד או , אם אשל , אלאם , , , אם אשל אַ אַ אאמעל (מאל או או איז איז איז איז איז איז איז א Ng, 、_DDA, 、Ngt Ngt Ng、_ , 凶肉、Ngt _ Mgg Mg,t / (、L IK1 $(\underline{M} = B1) \underline{M} \underline{M} \underline{M} \underline{J} \underline{t} \underline{M} \underline{M} \underline{t}$ B1 t, 1021018 _tt 088 . 01 IK1_104 B1, ___ 🕅 🕅 t、1023FC⊠、t,0232102_t(t_tt,_1K1) B1 🕅 t 🖉 🕅 🕅 .____ 🛯 📽 🖿 🕅 y_tt 🚬 t_t, IK1_⊠ 🕵 🖄 🙀 t. It , IK1 B1 🔤 t 🕻 🖉 ,_⊠ t . . . 1⊠-



Fig. 8. Genetic interactions between *SIK1* and *MOB1A*. (A) Ten-day-old *sik1–/- mob1a+/-* has more severe phenotypes than *sik1* alone, and *sik1–/mob1a–/-* is growth arrested. (B) The same phenotypes are observed on 4-week-old, soil-grown plants. (C) The 10-day-old transgenic line carrying *35S:MOB1A* in the *sik1-4* background has the same phenotypes as *sik1-4*. *35S:SIK1* can also not rescue *mob1a*. (D–G) Phenotypes consistent with (C) are observed in (D) 4-week-old, soil-grown plants; (E) 8-week-old plants; (F) mature siliques; (G) halves of siliques. Scale bars=1 cm in (A), (B) (by the double mutant), (C), (D), (F); 10 cm in (E); 1 mm in (G). (This figure is available in colour at *JXB* online.)

a, <u>N</u> N N N ฬ t 🕅 🖉 . I M D t_tt, odia a seekadaad , j ka a se se seka a seka ka ka 🌆 t (H. e a., 2010). H. 🖓 🖄 Ma 🕅 🖞 💆 👖 t, 🖓

 X
 X
 X
 X
 X
 X
 Y
 Y
 Y
 Y
 Y
 Y
 Y
 Y
 Y
 Y
 Y
 Y
 Y
 Y
 Y
 Y
 Y
 Y
 Y
 Y
 Y
 Y
 Y
 Y
 Y
 Y
 Y
 Y
 Y
 Y
 Y
 Y
 Y
 Y
 Y
 Y
 Y
 Y
 Y
 Y
 Y
 Y
 Y
 Y
 Y
 Y
 Y
 Y
 Y
 Y
 Y
 Y
 Y
 Y
 Y
 Y
 Y
 Y
 Y
 Y
 Y
 Y
 Y
 Y
 Y
 Y
 Y
 Y
 Y
 Y
 Y
 Y
 Y
 Y
 Y
 Y
 Y
 Y
 Y
 Y
 Y
 Y
 Y
 Y
 Y
 Y
 Y
 Y
 Y
 Y
 Y
 Y
 Y
 Y
 Y
 Y
 Y
 Y
 Y
 Y
 Y
 Y
 Y
 Y
 Y
 Y
 Y
 Y
 Y
 Y
 Y
 Y
 Y

Ng Nasut & Nasyat & Ang Ng K (1811 (1812) . 3). Ang Rayat K katalang ng t _XXXX (F. e a., 2009). A _ CDC42. XXXX _ 🖄 🕺 C IB 🖓 🛣 🖾 K1 , 🕰 t t, _tt, t. 6 IK1 🗤 🖾 🖄 🖾 🚬 (, _ _ _) 🖄 🖏 🖾 t t 🖄 , t, 🖄 🦯 لَّعَظَّ اتَهَ بِ _t _ لَفَ لِ _ CDC42 , _ E20 المُعَظَّ بِنَةٍ اللَّهُ اللَّهُ بِنَ مِعْظَ اللَّهُ ال tiona IKinanta nanka ta tian _a a a k ta ta ta . Na -1A/ , 46.2 (, 🔯 , ⊠ e . a., 2009) _⊠ _ I K11 (I , . . , 🖄 e . a., اُتھ کے آؤڈ کے اُور کے اُور کے اُس کے اُس کے اُس کے اُس کے اُس کے اُس کے اُس کے اُس کے اُس کے اُس کے اُس کے اُس 198812 - 図12181 第1989年2日、 (、y. e a., 2011), 1915日-1987日 - 1987日 - 19150-19150 boty bott both a have have a set to the set of the set kų Manganana karta na karta karta karta karta karta karta karta karta karta karta karta karta karta karta karta $\mathbf{M}_{\mathbf{M}} = \mathbf{M}_{\mathbf{M}} =$ X X X-لَاهَ ﴿ فَقَدْ هُوَ اللَّهِ اللَّهِ اللَّهِ اللَّهُ اللَّهُ اللَّهُ اللَّهُ اللَّهُ اللَّهُ اللَّهُ اللَّهُ ال $(\mathbf{y}, \mathbf{M}_{\mathbf{M}}, I, \mathbf{t}, \mathbf{t}) = 400 \,\mathbf{K}_{\mathbf{M}} \mathbf{M}_{\mathbf{M}} اُنھھ I (ھُر بھاھھ (اُنھھ G بِر مُجھ I) اُ 10月,1997年11月,1月1日,1997年11月,199 🕅 t. No No tà (2.42, -9), No , tà No ky ky ky Ήδ, 12_t, -, Ι, -, 152 <u>1</u>383 (JA) ⊠ t10833 – 🛛 🟦 🖾 t . 🛣 t 🕅 , - - C . IN ... t 1_N bla, _ JAZ $\begin{array}{c} \hline \begin{array}{c} \hline \end{array} \\ \\ \end{array} \\ \hline \end{array} \\ \\ \end{array} \\ \hline \end{array} \\ \\ \end{array} \\ \hline \end{array} \\ \\ \end{array} \\ \hline \end{array} \\ \\ \end{array} \\ \hline \end{array} \\ \\ \\ \end{array} \\ \\ \\ \end{array} \\ \\ \end{array} \\ \\ \end{array} \\ \\ \end{array} \\ \\ \\ \end{array} \\ \\ \\ \\ \end{array} \\ \\ \\ \end{array} \\ \\ \\ \\ \end{array} \\ \\ \\ \end{array} \\ \\ \\ \\ \end{array} \\ \\ \\ \end{array} \\ \\ \\ \\ \end{array} \\ \\ \\ \end{array} \\ \\ \\ \\ \end{array} \\ \\ \\ \end{array} \\ \\ \\ \end{array} \\ \\ \\ \\ \end{array} \\ \\ \\$ KM 题 t风 10, 100 100 10_ 10, 10, 10 (100, *e a*., 2007), 10, 1033, 、 tŽ_1⊠, Moz1a (a), t, JA _1≥/ C , _ , , 1≤ , . 1≤ t, , $- \boxtimes \mathbb{M}_{t,y} \boxtimes \mathbb{K}_{t,y} \land$ M t JANKK _t . t 🛛 IK1 (1880) ty _ÖXİMA _t 1880 JA 1880) TA 1880 A 18 1880 t. 🕅 _t. (.

Galla G, Zenoni S, Marconi G, et al. 2011. Sporophytic and gametophytic functions of the cell cycle-associated Mob1 gene in Arabidopsis thaliana L. Gene **484**, 1–12.

Goh HH, Sloan J, Dorca-Fornell J, Fleming A. 2012. Inducible repression of multiple expansin genes leads to growth suppression during leaf development. Plant Physiology **159**, 1759–1770.

Gonzalez N, Vanhaeren H, Inze D. 2012. Leaf size control: complex coordination of cell division and expansion. Trends in Plant Science **17**, 332–340.

Harris K, Lamson RE, Nelson B, Hughes TR, Marton MJ, Roberts CJ, Boone C, Pryciak PM. 2001. Role of scaffolds in MAP kinase pathway specificity revealed by custom design of pathway-dedicated signaling proteins. Current Biology **11**, 1815–1824.

Harvey K, Tapon N. 2007. The Salvador–Warts–Hippo pathway–an emerging tumour-suppressor network. Nature Reviews Cancer 7, 182–191.

Harvey KF, Pfleger CM, Hariharan IK. 2003. The Drosophila Mst ortholog, hippo, restricts growth and cell proliferation and promotes apoptosis. Cell **114**, 457–467.

Hepworth J, Lenhard M. 2014. Regulation of plant lateral-organ growth by modulating cell number and size. Current Opinion in Plant Biology **17**, 36–42.

Ho LL, Wei XM, Shimizu T, Lai ZC. 2010. Mob as tumor suppressor is activated at the cell membrane to control tissue growth and organ size in Drosophila. Developmental Biology **337**, 274–283.

Hofken T, Schiebel E. 2002. A role for cell polarity proteins in mitotic exit. EMBO Journal **21**, 4851–4862.

Hu Y, Poh HM, Chua NH. 2006. The Arabidopsis ARGOS-LIKE gene regulates cell expansion during organ growth. The Plant Journal **47**, 1–9.

Hu Y, Xie Q, Chua NH. 2003. The Arabidopsis auxin-inducible gene ARGOS controls lateral organ size. The Plant Cell **15**, 1951–1961.

Huang J, Wu S, Barrera J, Matthews K, Pan D. 2005. The Hippo signaling pathway coordinately regulates cell proliferation and apoptosis by inactivating Yorkie, the Drosophila homolog of YAP. Cell **122**, 421–434.

Ischebeck T, Stenzel I, Hempel F, Jin X, Mosblech A, Heilmann I. 2011. Phosphatidylinositol-4,5-bisphosphate influences Nt-Rac5-mediated cell expansion in pollen tubes of Nicotiana tabacum. The Plant Journal **65**, 453–468.

Jia J, Zhang W, Wang B, Trinko R, Jiang J. 2003. The Drosophila Ste20 family kinase dMST functions as a tumor suppressor by restricting cell proliferation and promoting apoptosis. Genes and Development **17**, 2514–2519.

Jonak C, Okresz L, Bogre L, Hirt H. 2002. Complexity, cross talk and integration of plant MAP kinase signalling. Current Opinion in Plant Biology 5, 415–424.

Karpov PA, Nadezhdina ES, Yemets AI, Matusov VG, Nyporko AY, Shashina NY, Blume YB. 2010. Bioinformatic search of plant microtubule- and cell cycle related serine-threonine protein kinases. BMC Genomics **11** Suppl 1, S14.

Krizek BA. 1999. Ectopic expression of AINTEGUMENTA in Arabidopsis plants results in increased growth of floral organs. Developmental Genetics **25**, 224–236.

Krizek BA. 2009. Making bigger plants: key regulators of final organ size. Current Opinion in Plant Biology **12**, 17–22.

Kurepa J, Wang S, Li Y, Zaitlin D, Pierce AJ, Smalle JA. 2009. Loss of 26S proteasome function leads to increased cell size and decreased cell number in Arabidopsis shoot organs. Plant Physiology **150**, 178–189.

Lamson RE, Winters MJ, Pryciak PM. 2002. Cdc42 regulation of kinase activity and signaling by the yeast p21-activated kinase Ste20. Molecular and Cellular Biology **22**, 2939–2951.

Leeuw T, Wu C, Schrag JD, Whiteway M, Thomas DY, Leberer E. 1998. Interaction of a G-protein beta-subunit with a conserved sequence in Ste20/PAK family protein kinases. Nature **391**, 191–195.

Li Y, Zheng L, Corke F, Smith C, Bevan MW. 2008. Control of final seed and organ size by the DA1 gene family in Arabidopsis thaliana. Genes and Development **22**, 1331–1336.

Lin M, Unden H, Jacquier N, Schneiter R, Just U, Hofken T. 2009. The Cdc42 effectors Ste20, Cla4, and Skm1 down-regulate the expression of genes involved in sterol uptake by a mitogen-activated protein kinase-independent pathway. Molecular Biology of the Cell **20,** 4826–4837.

Liu D, Gong Q, Ma Y, *et al*. 2010. cpSecA, a thylakoid protein translocase subunit, is essential for photosynthetic development in Arabidopsis. Journal of Experimental Botany **61**, 1655–1669.

Liu Y, Schiff M, Czymmek K, Talloczy Z, Levine B, Dinesh-Kumar SP. 2005. Autophagy regulates programmed cell death during the plant innate immune response. Cell **121**, 567–577.

Martin-Trillo M, Cubas P. 2010. TCP genes: a family snapshot ten years later. Trends in Plant Science 15, 31–39.

Massonnet C, Tisne S, Radziejwoski S, Vile D, De Veylder L, Dauzat M, Granier C. 2011. New insights into the control of endoreduplication: endoreduplication could be driven by organ growth in Arabidopsis leaves. Plant Physiology **157**, 2044–2055.

Menges M, de Jager SM, Gruissem W, Murray JA. 2005. Global analysis of the core cell cycle regulators of Arabidopsis identifies novel genes, reveals multiple and highly specific profiles of expression and provides a coherent model for plant cell cycle control. The Plant Journal **41**, 546–566.

Mizukami Y, Fischer RL. 2000. Plant organ size control: AINTEGUMENTA regulates growth and cell numbers during organogenesis. Proceedings of the National Academy of Sciences, USA **97**, 942–947.

Nelson BK, Cai X, Nebenfuhr A. 2007. A multicolored set of in vivo organelle markers for co-localization studies in Arabidopsis and other plants. The Plant Journal **51**, 1126–1136.

Nemoto K, Seto T, Takahashi H, Nozawa A, Seki M, Shinozaki M, Endo Y, Sawasaki T. 2011. Autophosphorylation profiling of Arabidopsis protein kinases using the cell-free system. Phytochemistry **72**, 1136–1144.

Omidbakhshfard MA, Proost S, Fujikura U, Mueller-Roeber B. 2015. Growth-regulating factors (GRFs): a small transcription factor family with important functions in plant biology. Molecular Plant **8**, 998–1010.

Pan D. 2010. The hippo signaling pathway in development and cancer. Developmental Cell **19**, 491–505.

Pantalacci S, Tapon N, Leopold P. 2003. The Salvador partner Hippo promotes apoptosis and cell-cycle exit in Drosophila. Nature Cell Biology **5**, 921–927.

Pinosa F, Begheldo M, Pasternak T, Zermiani M, Paponov IA, Dovzhenko A, Barcaccia G, Ruperti B, Palme K. 2013. The Arabidopsis thaliana Mob1A gene is required for organ growth and correct tissue patterning of the root tip. Annals of Botany **112**, 1803–1814.

Ponchon L, Dumas C, Kajava AV, Fesquet D, Padilla A. 2004. NMR solution structure of Mob1, a mitotic exit network protein and its interaction with an NDR kinase peptide. Journal of Molecular Biology **337**, 167–182.

Powell AE, Lenhard M. 2012. Control of organ size in plants. Current Biology **22,** R360–367.

Praskova M, Xia F, Avruch J. 2008. MOBKL1A/MOBKL1B phosphorylation by MST1 and MST2 inhibits cell proliferation. Current Biology **18**, 311–321.

Raitt DC, Posas F, Saito H. 2000. Yeast Cdc42 GTPase and Ste20 PAK-like kinase regulate Sho1-dependent activation of the Hog1 MAPK pathway. EMBO Journal **19**, 4623–4631.

Rawat SJ, Chernoff J. 2015. Regulation of mammalian Ste20 (Mst) kinases. Trends in Biochemical Sciences 40, 149–156.

Reyes FC, Buono R, Otegui MS. 2011. Plant endosomal trafficking pathways. Current Opinion in Plant Biology **14,** 666–673.

Roberts RL, Fink GR. 1994. Elements of a single MAP kinase cascade in Saccharomyces cerevisiae mediate two developmental programs in the same cell type: mating and invasive growth. Genes and Development **8**, 2974–2985.

Rock JM, Lim D, Stach L, et al. 2013. Activation of the yeast Hippo pathway by phosphorylation-dependent assembly of signaling complexes. Science **340**, 871–875.

Sablowski R, Carnier Dornelas M. 2014. Interplay between cell growth and cell cycle in plants. Journal of Experimental Botany **65**, 2703–2714.

Schnittger A, Weinl C, Bouyer D, Schobinger U, Hulskamp M. 2003. Misexpression of the cyclin-dependent kinase inhibitor ICK1/KRP1 in single-celled Arabidopsis trichomes reduces endoreduplication and cell size and induces cell death. The Plant Cell **15**, 303–315. Schruff MC, Spielman M, Tiwari S, Adams S, Fenby N, Scott RJ. 2006. The AUXIN RESPONSE FACTOR 2 gene of Arabidopsis links auxin signalling, cell division, and the size of seeds and other organs. Development **133**, 251–261.

Sessions A, Burke E, Presting G, et al. 2002. A high-throughput Arabidopsis reverse genetics system. The Plant Cell 14, 2985–2994.

Sheu YJ, Barral Y, Snyder M. 2000. Polarized growth controls cell shape and bipolar bud site selection in Saccharomyces cerevisiae. Molecular and Cellular Biology **20**, 5235–5247.

Soifer I, Barkai N. 2014. Systematic identification of cell size regulators in budding yeast. Molecular Systems Biology **10**, 761.

Sonoda Y, Sako K, Maki Y, Yamazaki N, Yamamoto H, Ikeda A, Yamaguchi J. 2009. Regulation of leaf organ size by the Arabidopsis RPT2a 19S proteasome subunit. The Plant Journal **60**, 68–78.

Spitzer C, Reyes FC, Buono R, Sliwinski MK, Haas TJ, Otegui MS. 2009. The ESCRT-related CHMP1A and B proteins mediate multivesicular body sorting of auxin carriers in Arabidopsis and are required for plant development. The Plant Cell **21**, 749–766.

Stavridi ES, Harris KG, Huyen Y, Bothos J, Verwoerd PM, Stayrook SE, Pavletich NP, Jeffrey PD, Luca FC. 2003. Crystal structure of a human Mob1 protein: toward understanding Mob-regulated cell cycle pathways. Structure **11**, 1163–1170.

Strange K, Denton J, Nehrke K. 2006. Ste20-type kinases: evolutionarily conserved regulators of ion transport and cell volume. Physiology **21**, 61–68.

Thines B, Katsir L, Melotto M, Niu Y, Mandaokar A, Liu G, Nomura K, He SY, Howe GA, Browse J. 2007. JAZ repressor proteins are targets of the SCF(COI1) complex during jasmonate signalling. Nature **448**, 661–665.

Tsukaya H. 2013. Leaf development. The Arabidopsis Book 11, e0163.

Udan RS, Kango-Singh M, Nolo R, Tao C, Halder G. 2003. Hippo promotes proliferation arrest and apoptosis in the Salvador/Warts pathway. Nature Cell Biology **5**, 914–920.

Uemura T, Kim H, Saito C, Ebine K, Ueda T, Schulze-Lefert P, Nakano A. 2012. Qa-SNAREs localized to the trans-Golgi network regulate multiple transport pathways and extracellular disease resistance in plants. Proceedings of the National Academy of Sciences, USA **109**, 1784–1789. Van Damme D, Bouget FY, Van Poucke K, Inze D, Geelen D. 2004. Molecular dissection of plant cytokinesis and phragmoplast structure: a survey of GFP-tagged proteins. The Plant Journal **40**, 386–398.

Weinl C, Marquardt S, Kuijt SJ, Nowack MK, Jakoby MJ, Hulskamp M, Schnittger A. 2005. Novel functions of plant cyclin-dependent kinase inhibitors, ICK1/KRP1, can act non-cell-autonomously and inhibit entry into mitosis. The Plant Cell **17**, 1704–1722.

Wu S, Huang J, Dong J, Pan D. 2003. hippo encodes a Ste-20 family protein kinase that restricts cell proliferation and promotes apoptosis in conjunction with salvador and warts. Cell **114**, 445–456.

Wuyts N, Dhondt S, Inze D. 2015. Measurement of plant growth in view of an integrative analysis of regulatory networks. Current Opinion in Plant Biology **25**, 90–97.

Xia T, Li N, Dumenil J, Li J, Kamenski A, Bevan MW, Gao F, Li Y. 2013. The ubiquitin receptor DA1 interacts with the E3 ubiquitin ligase DA2 to regulate seed and organ size in Arabidopsis. The Plant Cell **25**, 3347–3359.

Xia T, Xiao D, Liu D, Chai W, Gong Q, Wang NN. 2012. Heterologous expression of ATG8c from soybean confers tolerance to nitrogen deficiency and increases yield in Arabidopsis. PLoS One 7, e37217.

Xiong Y, McCormack M, Li L, Hall Q, Xiang C, Sheen J. 2013. Glucose–TOR signalling reprograms the transcriptome and activates meristems. Nature **496**, 181–186.

Yoshizumi T, Tsumoto Y, Takiguchi T, Nagata N, Yamamoto YY, Kawashima M, Ichikawa T, Nakazawa M, Yamamoto N, Matsui M. 2006. Increased level of polyploidy1, a conserved repressor of CYCLINA2 transcription, controls endoreduplication in Arabidopsis. The Plant Cell **18**, 2452–2468.

Yu FX, Guan KL. 2013. The Hippo pathway: regulators and regulations. Genes and Development **27**, 355–371.

Zenoni S, Fasoli M, Tornielli GB, Dal Santo S, Sanson A, de Groot P, Sordo S, Citterio S, Monti F, Pezzotti M. 2011. Overexpression of PhEXPA1 increases cell size, modifies cell wall polymer composition and affects the timing of axillary meristem development in Petunia hybrida. New Phytologist **191**, 662–677.

Zhao B, Li L, Lei Q, Guan KL. 2010. The Hippo–YAP pathway in organ size control and tumorigenesis: an updated version. Genes and Development **24**, 862–874.