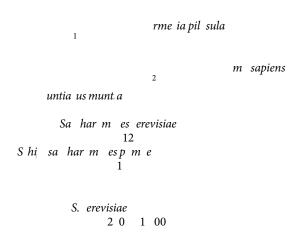
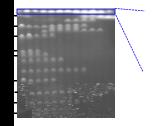
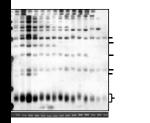
end-to-end chromosome fusions and centromere deletions. The fusion of sixteen native linear chromosomes into a single chromosome results in marked changes to the global three-dimensional structure of the chromosome due to the loss of all centromere-associated inter-chromosomal interactions, most telomere-associated inter-chromosomal interactions and 67.4% of intra-chromosomal interactions. However, the single-chromosome and wild-type yeast cells have nearly identical transcriptome and similar phenome profiles. The giant single chromosome can support cell life, although this strain shows reduced growth across environments, competitiveness, gamete production and viability. This synthetic biology study demonstrates an approach to exploration of eukaryote evolution with respect to chromosome structure and function.



Rationale

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## omosomal 3D structures

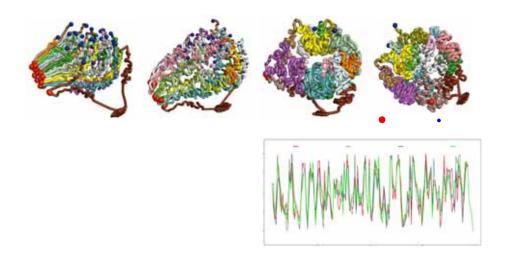
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2 S
Transcriptome and phenome analysis
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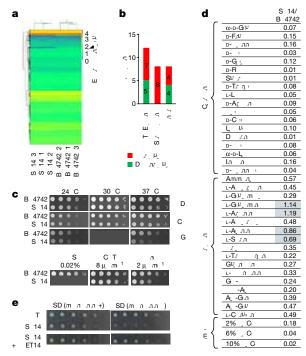


Fig. 4 Transcriptome and phenome analyses. a

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Meiosis and spore viability

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Online content

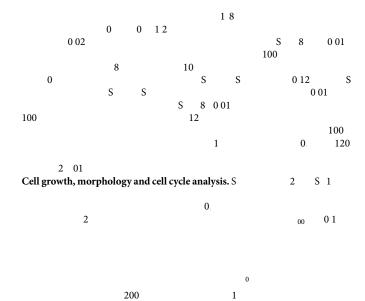
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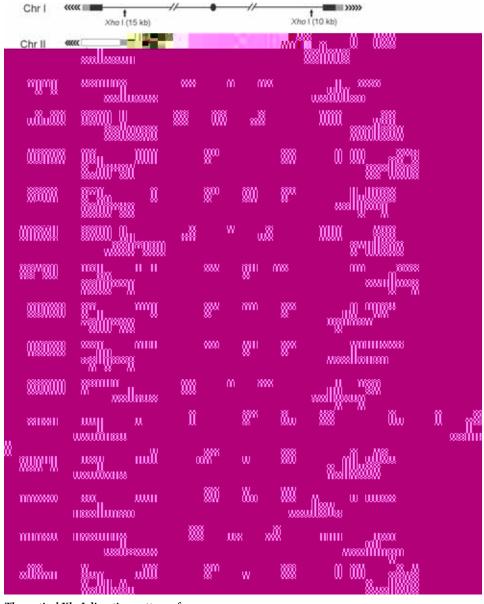
Plasmid constructions. 28

## **METHODS**

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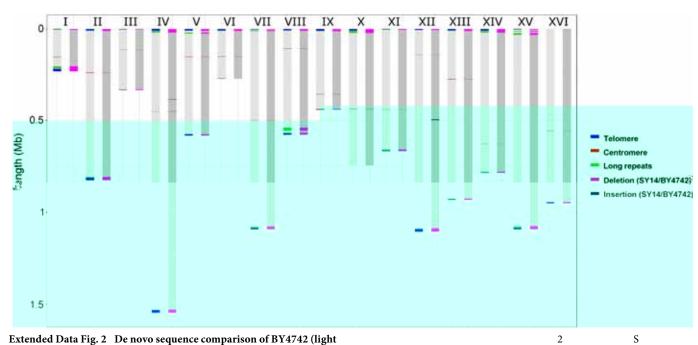


Extended Data Fig. 1 Theoretical XhoI digestion pattern of chromosome ends.

1

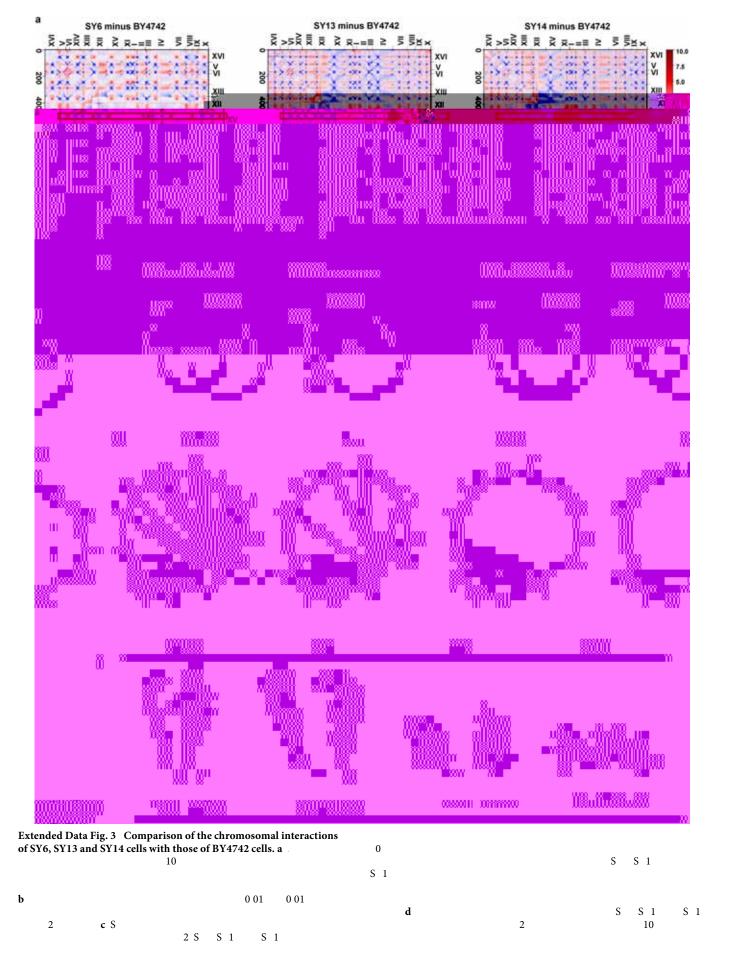
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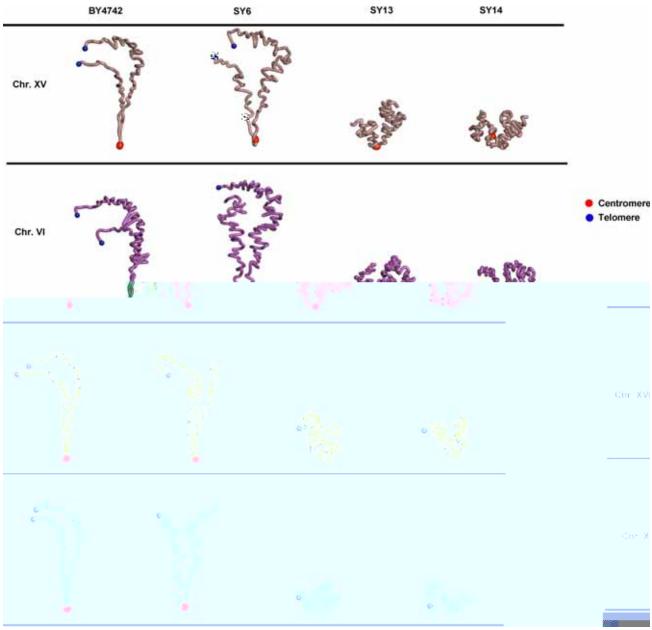
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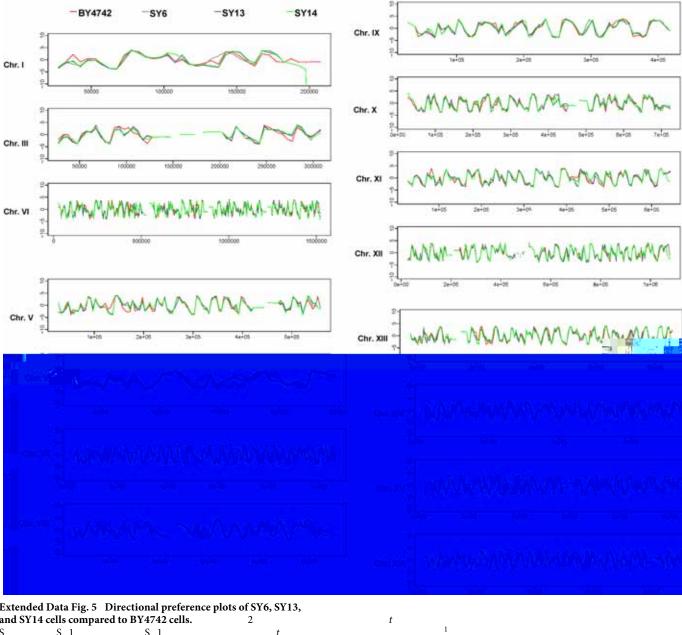
Extended Data Fig. 2  $\,$  De novo sequence comparison of BY4742 (light grey) and SY14 (dark grey) genomes.

2 S 1 S 1



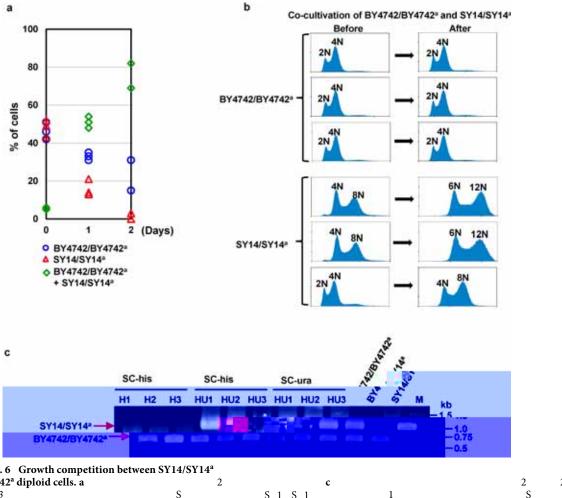


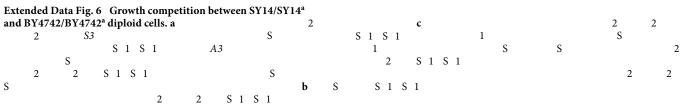
Extended Data Fig. 4 3D structures of single chromosomes.



Extended Data Fig. 5 Directional preference plots of SY6, SY13, and SY14 cells compared to BY4742 cells. 2 S 1 S 1

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#### E tended Data Table 1 | Details of the creation of a single chromosome | east

Strain	Chromosomes to fuse (chr. length in kb)	Newly fused chromosome (+) (chr. length in kb)	Newly deleted chromosome regions (RTel: right arm telomere sequence; LTel: left arm tel sequence) (Cen: centromere sequence; RS: repetitive sequence)	No. of transformants for chromosome fusion	Positive rate of chromosome fusion
SY0	VII (1090), VIII (560)	VII+VIII (1609)	VII: 5545-9584 (RS6), 1076068-1090940 (RTel, RS3); VIII: 524501-541100 (RS1), 1-8217 (LTel), 105447-106013 (Cen) V: 18067-23447 (RS11); XV: 10454-13126 (RS9), 21791-30776 (RS10,14);	100	1/3
SY1	XIII (920), XII (1080+1500)	XIII+XII (1980+1500)	XIII: 917281-924431 (RTel, RS7), 268013-268803 (Cen); XII: 1-14474 (LTel)	26	1/3
SY2	I (230), II (810)	I+II (1006)	l; 203183-230218 (RTel, RS1,2); ll: 1-9089 (LTel), 237784-238794 (Cen)	11	2/3
SY3	VI (270), XIV (780)	VI+XIV (1036)	VI: 269616-270161 (RTel), 148460-148774 (Cen); XIV: 1-17790 (LTel, RS4)	100	2/3
SY4	XVI (950), V (580)	XVI+V (1505)	XVI: 941976-948066 (RTel); V: 1-8079 (LTel), 151829-152588 (Cen)	143	4/8
SY5	IX (440), X (750)	IX+X (1160)	IX:436361-439888 (RTel), 355607-356006 (Cen); X:1-21750 (LTel, RS9,10,15)	94	1/4
SY6	III (320), IV (1530)	III+IV (1826)	III:313621-316620 (RTel), 114297-114969 (Cen); IV:1-19188 (LTel, RS12)	49	2/4
SY7	XVI-V (1505), VI-XIV (1036)	XVI-V+VI-XIV (2523)	V:569325-576874 (RTel); VI:1-8380 (LTel); XIV:628734-629219 (Cen)	130	1/5
SY8	XV (1090), XI (670)	XV+XI (1725)	XV:1073966-1091291 (RTel, RS8,7); XI: 439551-440264 (Cen), 1-3182 (LTel)	84	3/3
SY9	VII-VIII (1609), IX-X (1160)	VII-VIII+IX-X (2747)	VIII:552000-562643 (RTeI); IX:1-11214 (LTeI); X:436229-436425 (Cen)	30	2/5
QV10	I-II (1006), III-IV	I.HaIII.IV /2824\	1:151470-1517	5	2/4

The 'Strain' column lists the strain names, and the number in parentheses indicates the size of the native or fused chromosome in kilobase (kb). An orange plus indicates a fusion event; a dash between two chromosomes means that the fusion already occurred. 'Newly deleted chromosome regions' marks the deleted regions in the corresponding chromosomes; Rtel and Ltel in blue indicate the right arm and left arm of the corresponding telomere sequences, respectively; Cen in red indicates the corresponding centromere sequence; and RS represents repetitive sequences deleted in the corresponding chromosomes. The numbers for each region are referred from the S. cerevisiae S288C genome (http://www.yeastgenome.org/).



# E tended Data Table 2 $\mid$ Information regarding long repeat sequences near chromosome ends

Types of	Сору	Location on ch	romosomes (bp)
repeat sequences	number	RSs to be deleted	Retained RSs
RS1	3	I: 2034 5-219229; VIII: 525437- 539926	I:13089-27923
RS2	2	1:219230-229411	VIII: 539927-543610 549638- 556001
RS3	2	VII: 1076381- 1083886	II: 805133-812631
RS4	2	XIV: 7429-15942	VI: 5531-14039
RS5	2	XI: 658572-665429	III: 4327-11225
RS6	2	VII: 6223-9584	IX: 430983-434367
RS7	3	XIII: 917474- 923540; XV: 1078061- 1083736	XVI: 7409-13083
RS8	2	XV: 1073988- 1078544	XVI: 12601-17099
RS9	3	XV: 11053-13126; X: 8269-10330	IX: 8286-10347
RS10	3	XV:22397-27006; X: 16639-21229	IX:16656-21250
RS11	2	V: 18751-23447	XIV: 772693-777126
RS12	2	IV: 905-18681	X: 727164-744901
RS13	2	XII: 1059296-	III: 303903-308 <b>314</b>
荣814	2	XV: 27007-30776	IX: 21251-25254
RS15	2	X:10331-16638	DX: 10348-16655

This Table lists 15 types of long ( 2 kb) repeat sequences near telomeres, which have two or three copies. Only one copy of each long repeat sequence was retained, and the redundant copies were deleted in the SY14 strain. During the generation of the SY14 strain, six long repeats marked in red (that is, VII: 6223–9584 (RS6), VIII: 525437–539926 (RS1), V: 18751–23447 (RS11), XV: 11053–13126 (RS9), XV: 22397–27006 (RS10), and XV: 27007–30776 (RS14)), which are distal to telomeres, were deleted by two rounds of CRISPR–Cas9-mediated PCR targeting. The remaining 13 long repeats were deleted during chromosomal end-to-end fusions.



### E tended Data Table 3 | SNPs and indels confirmed by re-sequencing

Ref-			Mumin	a sequencing	Sange	r sequencing							
romosome	Ref-Loci	Ref base	wr	SY14	wr	SY14	Affected Gene	Variation Loci in Gene	Ref Codon	Variation Codon	Variation Type	Mutation in functional domain? (Y/N)	Null g
11	9136*	G	G	A	G	A	**	#1	+	100	-	-	
m	151645	G	G	A	G	A	-	75	.77	-7		7	77
x	517916	G	G	7	G	7	~	-	-		-	-7	-
xv	161691	c	c	G	c	G	-	23	2	12	12	-	-
1X	145512	G	G	1	G	T	Nup159	3198	CTC>L	CTA->L	synonymous	i.e.	-
#	447712	Α	Α.	τ.	A	τ.	S#2	4	AGTOS	TGT⇒C	non- synonymous	N	2
VR	775929	c	c	A	c	A	Skn1	737	GCT->A	GAT->D	non- synonymous	N	-
xv	454309	С	c	A	c	A	Vps5	542	ACA->T	ааа⇒к	non- synonymous	N	-
x	148653	G	G	A.	6	A	Yaki	1838	TCT⊹8	По€	non- synonymous	Y, Mutation within kinase domain	Null m grows in has inc fitnes lifesp sensit DNA do Null m
ХI	594128	с	c	Ą	c	A	Nup133	1304	TCT->S	тат-эч	non- synonymous	Y. Muta Story within WD40/YVTN repeat-like- containing domain	abnon elongat morphi decreas dea abno chem comp
XIV	225122	A	٨	G	A	G	Sqs1	1978	TGG⇔W	CGG⇒R	non- synonymous	Y. Mutation within R3H domain	Null m has dec or incr compe fitness on the condi
Ref- romosome	Ref-Loci	Ref base	ALTbase	Refilen	ALT len		equencing	Sanger sequ	_			lutation in	. 7
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a, SNPs. b, Indels.



### E tended Data Table 4 | Differentiall e pressed genes in SY14 compared to BY4742 cells

genelD	GeneSym	bol — groot		EU/12	Express		1109.5	1000 0	log-FC	Pvalue	FDR		Notes.	
	tjacent gener	311		5Y14-2	5Y14-3	WT-1	WT-2	WT-3	11.17940504	001110111	. (2000)		104075	
855848	ERR2	0.		0	0.04	2.08	1.52	2.05	-7.55	7.67E-13	6.78E-11	Chr XVI-L		
855849	HSP32			0.1	0	2.86	2.6	2.57	-6.83	2.83E-09	2.03E-07	Chr XVI-L		
855850	FEX2	1.	11	4.51	2.73	21.54	21.83	23.7	-3.03	1.64E-12	1.38E-10	Fourth gen	e near XVI-L	
855852	YPL277	C 13	97	2.82	2.09	7.11	5.57	8.61	-1.66	0.000071	0.004003	Sixth gene	near XVI-L.	
853625	мрнз	3.	79	7.58	7.24	19.12	18.21	19.61	-1.65	2.78E-05	0.001666	Second ger	ne near X-R.	
854634	VTH1	25	96	23.78	29.85	8.71	8.08	11.03	1.47	0.000126	0.006776	Second ger	te near IX-L, first gene was deleted in SY14.	
851230	SE01	6.	79	7.37	7.27	2.72	2.02	2.69	1.49	0.000254	0.013032	Third gene SY14.	near I-L, the first two genes were deleted in	
854002	YOL162	W 25	86	21.03	21.94	10,17	6.31	5.28	1.64	9.25E-05	0.005119	Fourth gen	e near XV-L.	
854001	YOL163			35.02	31.7	14.94	7.6	8.11	1.82	1.53E-05	0.000936		e near XV-L	
											1503335550	STATE OF THE PARTY	e near VI-L. first gene was deleted in SY14.	
850486	THIS	4.0	81	5.13	4.56	1.51	0.14	1.93	1,93	3.53E-05	0.002072		e near VII-R, first gene was deleted in S114.	
853207	MAL11	59	06	55.38	56.36	8.7	9,65	6.49	2.75	1.04E-11	8.32E	SY14.	e riear vii-rs, mat timee genes were deleted in	
850618	YFR057 response	W 22	58	20.31	18.16	3.13	1.46	0.96	3.40	1.64E-10	1.		ene near VI-R.	
8523		SP26	60.43	56.15	60.09	24.96	18.59	20.85	1,44	0.00026	0.013	364 Small	heat shock protein (sHSP) with chaperone a	
8547	1744 F	NR3	15,67	14.26	15.89	6.2	4.95	5.09	1.46	0.00018	0.005	714 DNA d	isoform of large subunit of ribonucleotide- sphate reductase; regulated by DNA replicati lamage checkpoint pathways, induced by DN je and replication stress.	
8533	1326 6	CA3	85.44	90.16	83.51	32.42	31	28.38	1,47	0.00014	0.007		n involved in mitochondrion organization; Ir SLN1-SKN7 osmotic stress signaling pathy	
8503	1331 F	BN1	30.17	22.64	28.11	10,71	6.16	8,97	1.63	9.49E-0	0.000	147 Simila increa	r to bacterial nitroreductases; protein abun ses in response to DN 100 Surformun	
	855932	OYES		01 6	.63 9.	95	1.71 2	2.57 2	1.83 1	1.69 7.0	BE-05	0.004003 III	eved NADPH oxido Ta se containing conomicleotide (FMN), has potential roles in tress response and programmed cell death	
	854944	HUGI	471	k.95 100	14.46 86	0.2	381.3 25	51.63 16	7.35 1	1.82 0.0	0048	0.023808 R	ibonucleotide reductase inhibitor; transcrip iduced by genotoxic stress and by activation ad\$1p pathway; protein abundance increase esponse to DNA replication stress. lasma membrane protein involved in mainta	
	850532	HSP12	21	1.2 22	3.37 18	1.79	62.8 5	14.47 46	L.85 1	1.98 1.5	5E-06	0.000102 0 8 1	embrane organization; involved in maintain ganization during stress conditions; induc- hock, oxidative stress, osmostress,; protein screased in response to DNA replication str	
	855132	HSP12				1.79					5E-06 6E-33	0.000102 0 8 10 3	sembrane organization; involved in maintain rganization during stress conditions; induce hock, exidative stress, comostress; protein creased in response to DNA replication stru- ubunit of telomeric Ku complex (Yku78p-Yk worked in telomere length maintanance, stress ossion effect, relocales to sit creame.	
ii le-	856132											0.000102 0 8 10 3	rembrane organization; involved in maintain riganization during stress conditions; induc- hock, oxidative stress, gemostress; protein creased in response to DNA replication stru- ubunit of telomeric Ku complex (YNu70p-YN wolved in telomere length maintaniance, str	
	855132 Others	YKUBO	12	.19 1	2.28 1	1.7	0 0	0.04 6	0.1 7	7.45 1.0	6E-33	0.000102 o 8 10 5 1.97E-31 te	pembrane organization; involved in maintain rganization during stress conditions; induc- tock, oxidative stress, osmostress, protein- creased in response to DNA replication str- ubunit of telomeric Ku complex (Yku76p-Yi	
1. 1. e-	855132 Others 916493	YKU80	12	.19 12 82.96	93,16	86,11	0 0	0.06 6	16,74	7.65 1.0 2.54	6E-33 1.53E-10	0.000102 0 8 8 8 1.97E-31 16 8	sembrane organization; involved in maintain rganization during stress conditions; induc- thock, oxidative stress, osmostress, protein- creased in response to DNA replication str- ubunit of telomeric Nu complex (YNA79-YI	
1	855132 Others 916493 85456	YKU80	12 137-1 T3	.19 1	93.16 230.81	86.11 526.59	13.39 265.15	0.06 ¢	0.1 7 16.74 154.28	7.65 1.0 2.54 1.25	1.53E-10 0.00102	0.000102 0 8 8 8 8 1.97E-21 to 9 1.13E-00 0.04968	sembrane organization; involved in maintal granization during stress conditions; induc- nock, oxidative stress, semostress, protein- creased in response to DNA replication stre- ubunit of telomeric Ku complex (Yku76y-Yi wolved in belomere length maintenance, stress of lossbon effect; relocates to sit stresses. Seavage to promote nonhomolo- during DSB repair.	
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1	Others 916493 85456 85229 85400 85185	YKU80  RDM FI YBR0 YOR2	12 137-1 13 12W-A 160W 61W-A	82.96 489.5 20.41 15.64 0.24	93.16 230.81 15.07 15.33	86.11 526.59 15.39 26.41	13.39 285.15 5.28 1.1 1.2	13.71 96.71 2.1 3.52 1.94	16.74 154.28 5.84 1.28	2.54 1.25 1.91 3.23 -3.57	1.53E-10 0.00102 2.22E-06 9.57E-07 3.33E-06	1.13E-00 0.000142 1.37E-31 1.13E-00 0.04968 0.00014: 6.39E-00	sembrane organization; involved in maintal riganization during stress conditions; industriant riganization during stress; conditions; protein controls, oxidative stress, osenostress; protein reasons to DNA replication stress; protein controls, and the control oxidative stress; consistent and controls to stress; controls to s	
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Sample size n 3. Exact negative binomial two-sided test was used to generate P values. Benjamini and Hochberg's algorithm was used to control the FDR.

# natureresearch

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